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GASTRO-PROTECTIVE ACTIVITY OF THE LEAVES AQUEOUS EXTRACT OF *DIOSPYROS MESPILIFORMIS* ON GASTRIC ULCERS IN SWISS MICE

Andre Perfusion Amang^{*1}, Parfait Bourvoune¹, Christophe Mezui², Gaël Tchokomeni Siwe³, Mesmine Teukam Kuissu⁴ and Paul Vernyuytan³

Department of Biological Sciences¹, Faculty of Science, University of Maroua, P.O. Box 814, Maroua, Cameroon.

Department of Biological Sciences², Higher Teachers' Training College, University of Yaoundé I. P. O. Box 47, Yaoundé, Cameroon.

Department of Animal Biology and Physiology³, Faculty of Science, University of Yaoundé I. P. O. Box 812, Yaoundé, Cameroon.

Institute of Medical Research and Study of Medicinal Plants⁴, P. O. Box 13033 Yaoundé, Cameroon.

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Correspondence to Author:

André Perfusion Amang

Department of Biological Sciences,
Faculty of Science, University of
Maroua, P. O. Box 814, Maroua,
Cameroon.

E-mail: perfusionamang@yahoo.fr

ABSTRACT: Information from ethnomedicine suggests that *Diospyros mespiliformis* possesses antiulcer properties. Thus, the purpose of this study was to evaluate its gastroprotective activity. The gastroprotective activity of the leaves aqueous extract of *D. mespiliformis* was evaluated by three experimental models of gastric ulcers in mice, namely: HCl/ethanol, HCl/ethanol with indomethacin pre-treatment and indomethacin (p. o). Ulcerated surface, ulcer index, and mucus mass were recorded. A qualitative phytochemical analysis of the extract was carried out. The administration of *D. mespiliformis* extract prevented the formation of gastric lesions against the necrotizing agents. For HCl/ethanol induction, the extract at 100, 200, and 400 mg/kg dose-dependently inhibited ulcer formation of 28.36, 29.19 and 35.82%, respectively. Indomethacin pre-treatment reduced the preventive effect of the extract to 19.69 and 28.24% at 200 and 400 mg/kg, respectively. As for indomethacin induction, the extract at 200 mg/kg showed the most important ulcer inhibition (88.13%). For all three induction models, a significant increase in mucus secretion between 44.75 and 121.34% was observed. *D. mespiliformis* could prevent the formation of gastric ulcers by stimulating mucus secretion *via* its direct action on mucus-secreting cells due to the presence of flavonoids, tannins, anthocyanins, and saponins in the extract.

INTRODUCTION: Gastric ulcer is a loss of substance from the gastric wall reaching deep into the muscularis¹ and is the most common disease of the gastrointestinal tract. It affects about 10% of the world population and particularly those in non-industrialized countries². It results from an imbalance between the aggressive and protective factors of the gastric mucosa³.

Aggressive factors may be exogenous, including *Helicobacter pylori* infection, alcohol, non-steroidal anti-inflammatory drugs, and tobacco consumption⁴ or endogenous, including hydrochloric acid, pepsin and reactive oxygen species (ROS)⁵. These factors often lead to a weakening of the protective factors, causing thus significant damage of the gastric mucosa, such as bleeding and especially inflammation⁶. Due to the multiple etiologies of gastric ulcers, several synthetic drugs are used for their treatment, namely: histamine H2 receptor antagonists, anticholinergics, proton pump inhibitors, antacids, prostaglandin analogs, and antibiotics⁷.



However, some of these drugs promote not only a wide range of adverse effects, but are also costly, making them inaccessible to most people in non-industrialized countries. So, phytotherapy seems to be a serious alternative for the management of gastric ulcers. For this purpose, several plants are used, namely: The roots of *Vernonia kotschyana*⁸. The leaves of *Eremomastax speciosa*⁹ the bark of the *Parkia biglobosa*¹⁰ and the bark, stems and roots of *Trichilia emetica*¹¹. *Diospyros mespiliformis* is a plant belonging to the Ebenaceae family. In Cameroon, it is listed among medicinal plants with a wide therapeutic arsenal, namely, antipyretic, analgesic, and anti-inflammatory effects¹². On the other hand, the healing activity of *D. mespiliformis* leaves on wounds has been cited in ethnomedicine by Burkill¹³, yet it is known that wound healing plants may have anti-ulcerogenic properties.

Thus, this study was designed to evaluate the gastro-protective activity of the leaves aqueous extract of *D. mespiliformis* on experimental models of gastric ulcers in mice.

MATERIALS AND METHODS:

Plant Material: The leaves of *Diospyros mespiliformis* were harvested in January 2018 in Mayo-Oulo in the North Region of Cameroon. They were collected with the help of a botanist (Professor Tchopsala). The plant has been authenticated at the Garoua Wildlife School by comparison with the existing specimen deposited under code HEFG/01404.

Animal Material: Male albino's mice, of 13 to 14 weeks old, with bodyweight ranging from 25 to 30 g were used. These mice were raised at the animal house of the University of Ngaoundéré (Cameroon). They were subjected to a natural day/night cycle. The diet was constituted of 50% maize flour, 20% soya flour, 10% cotton cake, 10% fish meal, 5% beef bone, 4% vegetable oil, 1% salt, with free access to tap water.

Preparation of the Extract: The leaves of *D. mespiliformis* after harvesting were washed and dried in a shed at room temperature. Using an electric shredder, the dry leaves were reduced in a fine powder and then weighed using a sensitive electrical scale. 250 g of this powder were

macerated in 2 liters of distilled water for 48 h. The macerate obtained was filtered using Whatman paper no. 3. The resulting filtrate was dried in an oven at a temperature of 40 °C for 24 h. The dry extract obtained had a mass of 10.5 g, with a yield of 4.2%, and was stored at 4 °C for subsequent tests.

Phytochemical Test: The leaves aqueous extract of *D. mespiliformis* was subjected to a qualitative phytochemical screening to detect the presence of some classes of phytoconstituents (phenols, alkaloids, flavonoids, terpenoids, sterols, tannins, glucoside, anthraquinone, coumarins, anthocyanins, saponins, and proteins) using the method described by Harborne¹⁴.

Gastric Ulcers Tests:

HCl / Ethanol-Induced Gastric Ulcers: The induction of gastric ulcers with HCl / ethanol was carried out according to the method described by Hara and Okabe¹⁵. 25 mice were deprived of food for 24 h before the experiment but with free access to water. These were divided into five groups of five animals, including three experimental groups which received the aqueous extract of *D. mespiliformis* orally (at the doses of 100, 200 and 400 mg/kg, respectively) and two control groups (positive and negative) which received orally, sucralfate (50 mg/kg) and distilled water (0.5 ml/ 30 g), respectively. After 1 h, all groups were given HCl/ethanol solution (150 mM/ 60%), and 1 h later, the mice were sacrificed. The stomachs were removed, and 5 ml of 2% formalin were injected into. The mucus from each stomach was gently scraped off by using glass slides, then the length and width of ulcers were measured, and scores assigned according to the method described by Tan et al.,¹⁶. The ulcer index (UI), percentage of inhibition (% I) and percentage of ulcerated surface (% US) were also calculated.

HCl/Ethanol-Induced Gastric Ulcers in Mice Pre-Treated with Indomethacin: HCl/ethanol-induced gastric ulcers with indomethacin pre-treatment were performed according to the model described by Sun et al.,¹⁷. The animals were divided into four groups of five mice each and fasted for 24 h before the experiment. They were given indomethacin intraperitoneally at the dose of 20 mg/kg. Thirty min later, the treated groups

received extract at the doses of 200 and 400 mg/kg. The positive and negative control groups received orally sucralfate (50 mg/kg) and distilled water (0.5 ml / 30 g), respectively. One hour later, all animals were given per os HCl/ethanol (0.5 ml / 30 mg). One hour later, these animals were sacrificed and the stomachs were examined in the same manner as for the HCl/ethanol model.

Indomethacin-Induced Gastric Ulcers: Gastric mucosa ulcers were induced by the method described by Pillai and Santhakumari¹⁸. After 24 h of fasting, the mice were divided into four groups of five mice each. The two test groups received extract (200 and 400 mg/kg, respectively) orally, while those in positive and negative control groups received sucralfate (50 mg/kg) and distilled water (0.5 ml / 30 g), respectively.

One hour later, all animals were given indomethacin solution per os at the dose of 50 mg/kg. 5 h later, the animals were sacrificed. The remaining procedure was the same as described for HCl / ethanol induction.

Statistical Analysis: The results obtained were expressed as mean \pm standard error on the mean (SEM). They were statistically analyzed using one-way ANOVA (analysis of variance), and multiple comparisons between groups were performed by the Student-Newman-Keuls post-test. The software Graph Pad Prism 5 was used for statistical analysis, and the values of $p < 0.05$ were considered as significant.

RESULTS:

Results of Phytochemical Test: Phytochemical analysis of the leaves aqueous extract of *D. mespiliformis* revealed the presence of the following classes of bioactive compounds: phenolic compounds, flavonoids, tannins, anthocyanins, saponins and proteins **Table 1**.

Antiulcer Tests:

Effects of the Leaves Aqueous Extract of *D. mespiliformis* on HCl / Ethanol-Induced Gastric Ulcers: Intragastric administration of HCl/ethanol to mice resulted in characteristic **Fig. 1** lesions on the glandular part of the stomach. These lesions are in the form of dark red bands, which are more numerous and larger size in the negative control **Fig. 1A**. Their size decreased by about half in the positive control **Fig. 1B**. The lesions size decreased more and gradually in dose-dependent (100, 200, and 400 mg/kg) manner in extract-treated groups, **Fig. 1C, 1D, and 1E**.

TABLE 1: RESULTS OF PHYTOCHEMICAL TEST OF THE LEAVES AQUEOUS EXTRACT OF *D. MESPILIFORMIS*

Classes of compounds	Observation
Phenolic compounds	+
Alkaloids	-
Flavonoids	+
Terpenoids	-
Tannins	+
Glucosides	-
Anthraquinones	-
Coumarins	-
Anthocyanins	+
Saponins	+
Proteins	+

+ = presence; - = absence

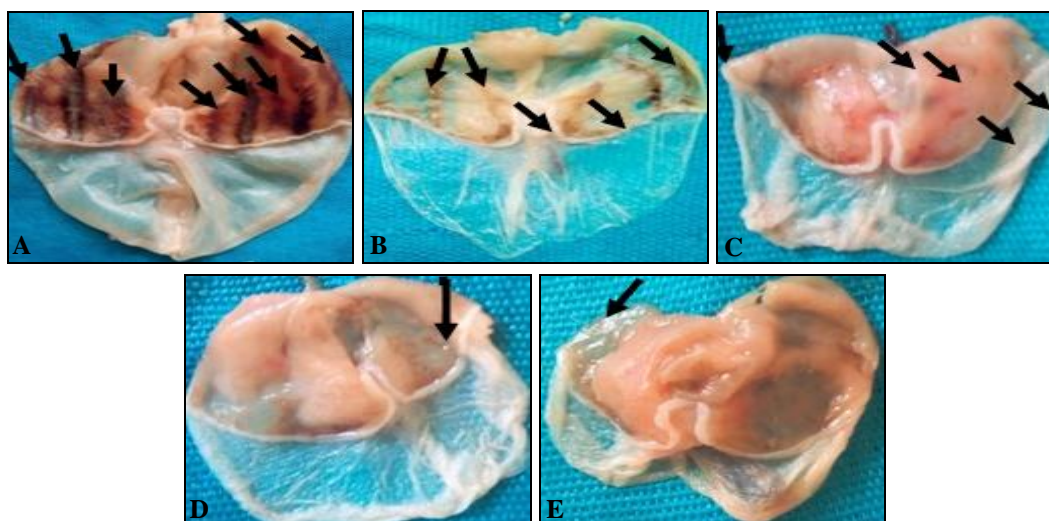


FIG. 1: PHOTOGRAPH OF STOMACHS ULCERATED WITH HCL/ETHANOL (a): Negative control, (b): Positive control, (c): 100 mg/kg of extract, (d): 200 mg/kg of extract, (e): 400 mg/kg extract dose: Indication of gastric ulcers

The leaves aqueous extract of *D. mespiliformis* (100, 200 and 400 mg/kg) induced a significant and dose-dependent decrease of the ulcerated area (12.60; 11.60 and 9.60 mm², respectively) compared to the negative control (42.40 mm²); corresponding to an increase in the inhibition

percentage of 28.36; 29.19 and 35.82, respectively. This inhibition of the degree of ulceration was accompanied by a significant increase of mucus secretion of 11.58; 12.34; and 16.82 mg, respectively, compared to the negative control (8.00 mg) **Table 2**.

TABLE 2: EFFECTS OF AQUEOUS EXTRACT OF *D. MESPILIFORMIS* ON HCl/ETHANOL-INDUCED GASTRIC ULCERS

Treatment	N	Dose (mg/kg)	US (mm ²)	% US	UI	% I	MM (mg)	% MM
Negative control	5	-	42.40 ± 4.28	13.25	4.83 ± 0.41	-	8.00 ± 1.938	
Sucralfate	5	50	24.60 ± 3.47**	8.31	3.77 ± 0.21***	21.95	9.88 ± 0.78	23.50
<i>D. mespiliformis</i>	5	100	12.60 ± 2.79***	3.66	3.46 ± 0.22***	28.36	11.58 ± 0.65	44.75
<i>D. mespiliformis</i>	5	200	11.60 ± 2.24***	3.65	3.42 ± 0.15***	29.19	12.34 ± 0.67	54.25
<i>D. mespiliformis</i>	5	400	9.60 ± 1.75***	3.00	3.10 ± 0.10***	35.82	16.82 ± 1.62**	110.25

The values are expressed as mean ± SEM; ** p<0.01; *** p<0.001; significant differences compared to the negative control using student-newman-keuls multiple comparison test followed by one-way anova; n = 5, number of mice per groups; us = ulcerated surface; % us = percentage of ulcerated surface; ui = ulcer index; %i = percentage of inhibition; mm = mucus mass; % mm = percentage mucus increase

Effects of the Leaves Aqueous Extract of *D. Mespiliformis* on HCl / Ethanol-Induced Gastric Ulcers with Indomethacin Pre-treatment: Macroscopic observations showed, in the glandular part of the stomach, formation of gastric lesions

induced by HCl/ethanol with indomethacin pre-treatment. The lesions were larger and more numerous in the negative control, and they decreased in animals treated with the extract and sucralfate **Fig. 2**.

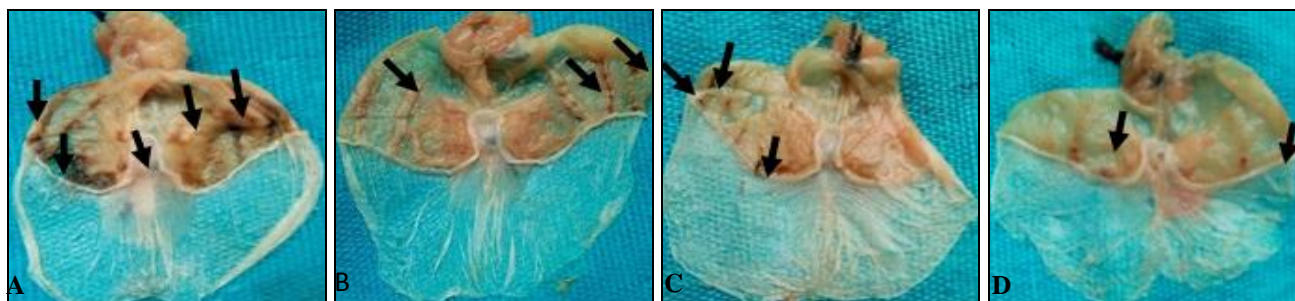


FIG. 2: PHOTOGRAPHS OF STOMACHS ULCERATED WITH HCL/ETHANOL WITH INDOMETHACIN PRETREATMENT (a) Negative control; (b) Positive control; (c) Dose 200 mg/kg extract; (d) Dose 400 mg/kg extract, :Indication of gastric ulcers

Indomethacin pre-treatment reduced the preventive effects of the extract (200 and 400 mg/kg), evidenced by a decrease of the percentage of inhibition for HCl/ethanol induction with indomethacin pre-treatment (19.69 and 28.65, respectively) **Table 3**, compared to HCl/ethanol

induction (29.19 and 35.82, respectively) **Table 2**. This reduction in preventive effects was accompanied by a decrease in mucus secretion at the dose of 400 mg/kg of extract from 16.82 mg to 10.46 mg after pre-treatment with indomethacin **Table 2** and **3**.

TABLE 3: EFFECTS OF AQUEOUS EXTRACT OF *D. MESPILIFORMIS* ON HCL/ETHANOL-INDUCED GASTRIC ULCERS WITH INDOMETHACIN PRETREATMENT

Treatment	N	Dose (mg/kg)	US (mm ²)	% US	UI	% I	MM (mg)	% MM
Negative control	5	-	23.60 ± 3.17	7.38	3.86 ± 0.18		4.78 ± 0.68	
Sucralfate	5	50	28.80 ± 3.80	9.00	3.46 ± 0.13	10.36	9.20 ± 0.18***	92.47
<i>D. mespiliformis</i>	5	200	10.60 ± 1.90*	3.31	3.10 ± 0.10	19.69	10.58 ± 1.93***	121.34
<i>D. mespiliformis</i>	5	400	8.60 ± 2.36*	2.69	2.77 ± 0.69*	28.24	10.46 ± 1.81***	118.83

The values are expressed as Mean ± SEM; * p < 0.05; *** p < 0.001; significant differences compared to the negative control using student-Newman-Keuls multiple comparison test followed by one-way ANOVA; N = 5, number of mice per groups; US = ulcerated surface; % US=percentage of ulcerated surface; UI = ulcer index; %I = percentage of inhibition; MM = mucus mass; % MM = percentage of mucus increase

Effects of the Leaves Aqueous Extract of *D. Mespiliformis* on Indomethacin-Induced Gastric Ulcers: Extract at the dose of 200 mg/kg prevented ulcers formation (88.13%) induced with indomethacin, corresponding to a significant

decrease of ulcerated surface (0.40 mm^2) compared to negative control (11.40 mm^2) **Fig. 3.** Extract (200 mg/kg) induced also a significant increase ($p < 0.001$) of mucus production (10.90 mg) compared to negative control (5.50 mg) **Table 4.**

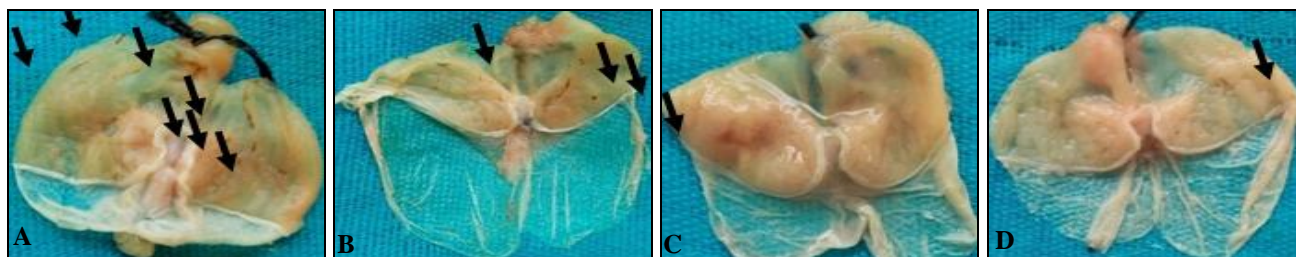


FIG. 3: PHOTOGRAPH OF STOMACHS ULCERATED WITH INDOMETHACIN (a) Negative control; (b) Positive control; (c) Dose 200 mg/kg extract; (d) Dose 400 mg/kg extract, indication of gastric ulcers

TABLE 4: EFFECTS OF AQUEOUS EXTRACT OF *D. MESPILIFORMIS* ON GASTRIC ULCERS INDOMETHACIN- INDUCED

Treatment	N	Dose (mg/kg)	US	% US	UI	% I	MM	% MM
Négative control	5	-	11.40 ± 3.37	3.56	3.37 ± 0.30		5.50 ± 0.53	-
Sucralfate	5	50	$3.00 \pm 1.38^{**}$	0.94	$1.80 \pm 0.73^*$	46.59	6.26 ± 0.79	13.81%
<i>D. mespiliformis</i>	5	200	$0.40 \pm 0.40^{***}$	0.13	$0.40 \pm 0.40^{**}$	88.13	$10.90 \pm 1.19^{**}$	98.18%
<i>D. mespiliformis</i>	5	400	$0.80 \pm 0.80^{***}$	0.25	$0.60 \pm 0.60^{**}$	82.20	$9.50 \pm 1.83^*$	80.00%

The values are expressed as mean \pm SEM; ** $p < 0.05$; * $p < 0.01$; *** $p < 0.001$; significant differences compared to the negative control using student-newman-keuls multiple comparison test followed by one-way ANOVA; N = 5, number of mice per groups; US= ulcerated surface % US = percentage of ulcerated surface UI = ulcer index; % I = percentage of inhibition; MM = mucus mass; % MM = percentage of mucus increase

DISCUSSION: *Diospyros mespiliformis* possesses a wide range of medicinal uses¹⁹. Based on its numerous virtues, the study of its gastroprotective properties was investigated by using three models of gastric ulcer induction: HCl/ethanol induction, HCl/ethanol induction with indomethacin pre-treatment and indomethacin induction in mice.

Several factors, both endogenous and exogenous, are responsible for the establishment of peptic ulcers, following an imbalance between aggressive and protective factors of the gastric mucosa. Alcohol consumption and NSAID intake are among the most common and dangerous ulcerogenic factors leading to gastroduodenal damage²⁰. For this reason, models of ulcer lesions with HCl/ethanol and indomethacin were selected. Indomethacin is the first choice of NSAID for producing the experimental ulcers because it has the highest ulcerogenic potential compared to other NSAIDs²¹. Macroscopic observations of stomachs subjected to HCl/ethanol showed characteristic gastric lesions in the glandular part of the stomach; these are in the form of dark red bands. These lesions are characteristic of the lesions induced by this mixture, similarly to the observations of studies

by Anandan *et al.*,²² Silva *et al.*,²³ and Martins *et al.*,²⁴ which showed the necrotic power of HCl/ethanol on the stomachs of rats. The HCl/ethanol mixture acts by exerting a direct toxic effect on the epithelium, leading to the formation of characteristic necrotic lesions due to a vasoconstrictor effect on veins and arteries of the gastric mucosa, to a decrease in blood flow thus producing congestion and inflammation²⁵ solubilization of stomach mucus components and oxidative stress²⁴.

The results of this study show that the leaves aqueous extract of *D. mespiliformis* at 200 and 400 mg/kg significantly ($p < 0,001$) reduced the percentage of ulceration (3.65 and 3.00) corresponding to percentages of inhibition of 29.19 and 35.82, respectively. These results suggest that the extract protects the gastric mucosa against the ulcerogenic agent similarly to by Antonio *et al.*,²⁶ who showed that the *Solanum variable* extract prevented the formation of gastric lesions induced by HCl / ethanol. The leaves aqueous extract of *D. mespiliformis* (100, 200, and 400 mg/kg) induced a very significant ($p < 0.01$) increase of mucus secretion (11.58, 12.34 and 16.8 mg, respectively)

compared to the negative control (8 mg). Indeed, mucus constitutes a line of defense as mentioned by Pasquier²⁷; it is characterized by a film formed by the polymerization of glycoproteins that traps bicarbonates, to delay the penetration of endoluminous H⁺ ions and thus establish a pH gradient ranging from less than 3 at the luminal surface of this layer, to more than 7 on the mucous surface.

To determine the cytoprotective mechanism of action by which the leaves aqueous extract of *D. mespiliformis* acts to increase mucus secretion, the HCl/ethanol induction with indomethacin pre-treatment was performed. Mice pre-treated with indomethacin showed ulcers similar to those obtained with HCl / ethanol. Indomethacin is a non-steroidal anti-inflammatory drug (NSAID) that inhibits the secretion of endogenous prostaglandins. These prostaglandins protect the stomach against damage by stimulating bicarbonate and mucus secretion, maintaining gastric microcirculation, and regulating stomach mucosal repair^{28, 29}.

The inhibitory action of indomethacin exposes the gastric mucosa, thus allowing the HCl/ethanol mixture to attack the gastric wall and generate ulcers. This would explain the severity of the lesions observed on the mucosa of mice subjected to pre-treatment with indomethacin. A decrease in the ulcerated surface was observed in the extract-treated groups at the doses of 200 (10.60 mm²) and 400 mg/kg (8.60 mm²) compared to the negative control (23.60 mm²).

The extract (200 and 400 mg/kg) also increased mucus secretion (10.58 and 10.46 mg, respectively) compared to the negative control group (4.78 mg). These results are in line with those obtained by Mezui et al.,³⁰ who had shown that the increase in mucus secretion by the aqueous extract of *Cassia arereh* following HCl/ethanol with indomethacin pre-treatment model was not related at the action of endogenous prostaglandins; which could be the case of our extract. To verify the gastro-protective mechanism of action of the leaves aqueous extract of *D. mespiliformis*, the induction method with indomethacin was performed. The main mechanism by which indomethacin induces gastric ulcers implies inhibition of the biosynthesis of endogenous prostaglandins by inhibiting cyclooxygenase.

Indeed, indomethacin causes non-selective inhibition of cyclo-oxygenase 1 and cyclo-oxygenase 2, which are enzymes responsible of the synthesis of prostaglandins. Prostaglandins play an important cytoprotective role in the gastric mucosa by positively influencing mucus and bicarbonate secretion, surface epithelial cells and mucosal circulation^{31, 32}. Their absence leads to mucosal damage resulting in gastric ulcers^{33, 34}. Analysis of our results shows that treatment of mice with aqueous extract of *D. mespiliformis* leaves (200 and 400 mg/kg) significantly inhibited ulcer formation (88.13 and 82.20%) associated with a significant increase in mucus secretion (10.90 and 9.50 mg) compared to the negative control (5.50 mg). These results suggest that the extract may have a direct action on mucus-secreting cells.

The observed anti-ulcer activity of the extract could be attributed to its richness in classes of bioactive compounds that would strengthen the mucobicarbonate barrier. Phytochemical tests revealed the presence of several classes of bioactive compounds such as flavonoids, tannins, saponins and anthocyanins. These results are similar to those obtained by Adeniyi et al.,³⁵ as classes of bioactive compounds revealed by the qualitative phytochemical analysis were the same. Several studies have demonstrated the anti-ulcerogenic effect of tannin-rich plants. Gege-Adebayo et al.,³⁶ associated the anti-ulcerogenic potential of *Ocimum gratissimum* with its high tannin content. Indeed, tannins prevent the formation of ulcers due to their ability to precipitate proteins forming a protective layer, thus, preventing the action of aggressive factors on the gastric mucosa.

Their vasoconstrictive and astringent action on the site of the ulcer forms an impermeable layer on the wall and thus prevents the formation of gastric ulcers³⁷. As for the flavonoids, several mechanisms have been proposed to explain their gastroprotective effects, including enhancement of prostaglandin secretion in the gastric mucosa and decrease of histamine secretion by mast cells through inhibition of histidine decarboxylase³⁸.

This anti-ulcer activity might also be due to anthocyanins activity, as Alvarez-Suarez et al.,³⁹ have shown that anthocyanins increase GSH levels and antioxidant enzyme activity. For saponins, their

protective activities are linked to the fact that they stimulate mucus production factors (prostaglandins) in the mucosa.

CONCLUSION: At the end of this work, it appears that the leaves aqueous extract of *Diospyros mespiliformis* protected the gastric mucosa against the damage induced by various necrotizing agents. The gastro-protective action could be attributed to the increase in mucus secretion by a mechanism that is not linked to the action of endogenous prostaglandins but rather to the direct stimulation of mucus secretory cells, due of the presence of flavonoids, tannins, anthocyanins and saponins in this extract.

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CONFLICTS OF INTEREST: Authors declare no conflict of interest.

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