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## CYTOTOXIC EFFECT OF *ALPINIA NIGRA* CRUDE EXTRACTS OBTAINED FROM ITS RHIZOMES

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### Keywords:

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**ABSTRACT:** The present study was aimed to investigate the cytotoxic activity of different fractions (petroleum ether, ethyl acetate & chloroform) of the rhizome of *Alpinia nigra* – a plant belonging to the family Zingiberaceae. The rhizome of *Alpinia nigra* was extracted with organic solvent, and the extracts were fractionated by using three fractions (petroleum ether, ethyl acetate & chloroform). The petroleum ether, ethyl acetate & chloroform soluble fractions of *Alpinia nigra* were screened for cytotoxic activity using brine shrimp lethality bioassay. A reputed cytotoxic agent vincristine sulphate was used as a positive control. From the results of the brine shrimp lethality bioassay it can be well predicted that petroleum ether, ethyl acetate and chloroform-soluble fractions of *Alpinia nigra* rhizome possess cytotoxic principles (with LC<sub>50</sub> 1.245 µg/ml and LC<sub>50</sub> 2.151 µg/ml and LC<sub>50</sub> 2.737 µg/ml respectively) comparison with positive control vincristine sulphate (with LC<sub>50</sub> 0.563 µg/ml). All the extracts of *Alpinia nigra* showed moderate cytotoxic activities in this experiment. The results of this study are highly promising.

**INTRODUCTION:** Plants are one of the most important sources of medicines. Today a large number of drugs in use is derived from plants. The uses of medicinal plants as traditional medicines are well known in rural areas of Bangladesh. Our present study was designed to determine the cytotoxic activity of different fractions (petroleum ether, ethyl acetate & chloroform) of the rhizome of *Alpinia nigra*. *Alpinia nigra* is a perennial rhizomatous herb with tuberous, aromatic roots. Leaves are sessile, oblong, lanceolate, acuminate, cuspidate, glabrate, striate and compressed.

Flowers are inodorous, pink, inerect, arranged in dense panicles with a pubescent rachis and small ovate cupular bracts.

Fruits are black, thin, globose and irregularly rupturing. Seeds are many, small, black and angular <sup>1</sup>. In *Alpinia nigra*, flowers are pink <sup>2</sup>. An anti-microbial diterpene has been isolated from the rhizome <sup>3</sup>. The rhizomes are bitter, acrid, thermogenic, aromatic, nervine tonic, stimulant, revulsive, carminative, stomachic, disinfectant, aphrodisiac, expectorant, broncho-dilator, anti-fungal, febrifuge, anti-inflammatory and tonic <sup>4</sup>. Rhizome is CVS and CNS active, diuretic, hypothermic. Seed is anti-ulcerative <sup>5</sup>. Alcohol (50%) extract of the rhizome is anti-amphetaminic. Unani physicians consider it good for impotence <sup>6</sup>. The volatile steam oil stimulates bronchial glands when exposed to its vapors <sup>7</sup>. The rhizomes of *Alpinia nigra* are anti-inflammatory <sup>8</sup>.



The pharmacognosy and toxicology of anti-carcinogenic natural products from galanga root oil have been studied by Zheng<sup>9</sup>. It has been found that oral administration of ethanolic extract of the rhizome produced a hypolipidemic effect by reducing serum and tissue levels of total cholesterol, triglycerides and phospholipid in Wister rats<sup>10</sup>.

## MATERIAL AND METHODS:

**Selection of Plant:** The fresh rhizome of the plant *Alpinia nigra* was selected for observation of cytotoxic activities. The fresh rhizomes of *Alpinia nigra* were collected during June 2010 from the area of Gazipur.

**Drying and Pulverization:** The fresh rhizomes of the plants were first washed with water to remove adhering dirt and then cut into small pieces, sun-dried for 4 days and finally dried at 45 °C for 36 h in an electric oven. After complete drying, the entire portions were pulverized into a coarse powder with the help of a grinding machine and were stored in an airtight container for further use.

**Cold Extraction of Plant Material:** The fresh rhizome of *Alpinia nigra* was collected, dried and ground to a coarse powder. The powder sample (500 gm) was subjected to cold extraction with methanol for about 30 days. The methanol extract was then subjected to the modified Kupchan partitioning method with pet ether, ethyl acetate, and chloroform. Thus, three extractives were obtained, leaving the aqueous fraction.

**Preparation of Mother Solution:** 5 gm of methanol extract of *Alpinia nigra* was triturated with 90 ml of methanol containing 10 ml of distilled water. The crude extract went to the solution completely. This is the mother solution, which was partitioned off successively by three solvents of different polarity.

**Partitioning with Pet Ether:** The concentrated methanol extract was made the slurry with water. The crude extract is diluted with sufficient amount of aqueous methanol (90%) and then gently shaken in a separating funnel with 100ml of a suitable organic solvent (such as pet ether) which is immiscible with aqueous methanol. The mixture is kept undisturbed for several minutes for separation of the organic layer from the aqueous phase. The

materials of the crude extract will be partitioned between the two phases depending on their affinity for the respective solvents. The organic layer is separated, and this process is carried out thrice for maximum extraction of the samples. The pet ether layer (upper layer) was collected. The process was repeated three times. The combined pet ether extract was concentrated. After separating off the organic phase, the aqueous phase thus obtained is successively extracted with other organic solvents, usually of the increasing polarity (such as ethyl acetate, chloroform, etc.). Finally, all the fractions (organic phases as well as the aqueous phase) are collected separately and evaporated to dryness. These fractions are used for isolation of compounds.

**Partitioning with Ethyl Acetate:** After pet ether extraction, 10 ml of distilled water was added and mixed. Then ethyl acetate (100 ml) was added to the mother solution, and the mass was shaken vigorously in a separating funnel. Then the funnel was allowed to stand for a few minutes for the complete separation of the layers. The ethyl acetate (organic lower layer) fractions were collected together and evaporated. The process was repeated two times. The aqueous fraction was preserved for the next step.

**Partitioning with Chloroform:** After ethyl acetate extraction, 16 ml of distilled water was added and mixed uniformly. Then, chloroform (100 ml) was added to the mother solution, and the mass was shaken vigorously in a separating funnel. Then the funnel was allowed to stand for a few minutes for the complete separation of the layers. The chloroform (organic lower layer) fractions were collected together and evaporated. The process was repeated three times. The aqueous methanolic fraction was preserved as an aqueous fraction.

**Cytotoxicity Test:** Pharmacology is simply toxicology at a lower dose, and toxicology is simply pharmacology at a higher dose. Bioactive compounds are almost always toxic in high doses. The *in-vivo* lethality in a simple zoologic organism can be used as a convenient monitor for screening and fractionation in the discovery and monitoring of bioactive natural products. It focused on *Artemia salina* as a test organism and developed a protocol for Brine shrimp lethality bioassay to monitor cytotoxicity of a compound.

**Brine Shrimp Lethality Bioassay:**

**Principle:** Brine shrimp eggs are hatched in stimulated sea water to get nauplii. Test samples are prepared by dissolving in DMSO and by addition of the calculated amount of DMSO, the desired concentration of the test sample is prepared. Ten nauplii are counted by visual inspection and are taken in vials containing 5 ml of simulated sea water. Then samples of different concentration are added to the premarked vials through a micropipette. The vials are then left for 24 h, and then the nauplii are counted again to find out the cytotoxicity of the test agents. These data are processed in a simple program to estimate LD<sub>50</sub>

values with 95% confidence intervals for statistically significant comparisons of potencies.

**Materials:**

- *Artemia salina* leach (brine shrimp eggs)
- Sea salt (NaCl)
- A small tank with a perforated dividing dam to hatch the shrimp
- Lamp to attract shrimps
- Pipettes (5, 25 ml) and Micropipette (5-40 $\mu$ l)
- Glass vials
- Magnifying glass
- Test samples of experimental plants

**TABLE 1: TEST SAMPLES OF EXPERIMENTAL PLANTS**

Plant	Test samples	Measured Amount (mg)
<i>Alpinia nigra</i>	Pet ether fractions of the methanolic extract	4.00
	Ethyl acetate fractions of the methanolic extract	4.00
	Chloroform fractions of the methanolic extract	4.00

**Procedure:**

**Preparation of Seawater:** 38 gm sea salt (pure NaCl) was weighed, dissolved in one liter of distilled water and filtered off to get a clear solution.

**Hatching of Brine Shrimps:** *Artemia salina* leach (brine shrimp eggs) collected from pet shops was used as the test organism. Seawater was taken in the small tank, and shrimp eggs were added to one side of the tank, and then this side was covered. Two days were allowed to hatch the shrimp and to be matured as nauplii. Constant oxygen supply was provided throughout the hatching time. The hatched shrimps were attracted to the lamp through the perforated dam and with the help of a Pasteur pipette 10 living shrimps were added to each of the vials containing 5 ml of seawater.

**Preparation of Test Solutions with Samples of Experimental Plants:** Clean test tubes were taken. These test tubes were used for eight different concentrations (one test tube for each concentration) of test samples, and eight test tubes were taken for standard drug Vincristine for eight concentration of it and another one test tubes for the control test. Then 100  $\mu$ l of the solution was taken in test tube each containing 5 ml of simulated seawater and 10 shrimp nauplii. Thus, the final concentration of the prepared solution in the first test tube was 400  $\mu$ g/ml.

Then a series of solutions of varying concentrations were prepared from the stock solution by serial dilution method. In each case 100  $\mu$ l sample was added to test tube and fresh 100  $\mu$ l DMSO was added to the vial. Thus the concentrations of the obtained solution in each test tube were as- 400  $\mu$ g/ml, 200  $\mu$ g/ml, 100  $\mu$ g/ml, 50  $\mu$ g/ml, 25  $\mu$ g/ml, 12.5  $\mu$ g/ml, 6.25  $\mu$ g/ml, 3.125  $\mu$ g/ml, 1.5625  $\mu$ g/ml 0.7815  $\mu$ g/ml.

**Preparation of Control Group:** Control groups are used in cytotoxicity study to validate the test method and ensure that the results obtained are only due to the activity of the test agent and the effects of the other possible factors are nullified. Usually, two types of control groups are used as i) Positive control ii) Negative control.

**Preparation of the Positive Control Group:** Positive control in a cytotoxicity study is a widely accepted cytotoxic agent, and the result of the test agent is compared with the result obtained for the positive control. In the present study, vincristine sulphate is used as the positive control. Measured amount of the vincristine sulphate is dissolved in DMSO to get an initial concentration of 20  $\mu$ g/ml from which serial dilutions are made using DMSO to get 10  $\mu$ g/ml, 5  $\mu$ g/ml, 2.5  $\mu$ g/ml, 1.25  $\mu$ g/ml, 0.625  $\mu$ g/ml, 0.3125  $\mu$ g/ml and 0.15625  $\mu$ g/ml. Then the positive control solutions are added to the premarked vials containing ten living brine shrimp

nauplii in 5 ml simulated sea water to get the positive control groups.

**Preparation of the Negative Control Group:** 100  $\mu$ l of DMSO was added to each of three premarked glass vials containing 5 ml of simulated seawater and 10 shrimp nauplii to use as control groups. If the brine shrimps in these vials show a rapid mortality rate, then the test is considered as invalid as the nauplii died due to some reason other than the cytotoxicity of the compounds.

**Counting of Nauplii:** After 24 h, the vials were inspected using a magnifying glass, and the number of survived nauplii in each vial was counted. From this data, the percent (%) of lethality of the brine shrimp nauplii was calculated for each concentration.

**Result and Discussion of Brine Shrimp Lethality Bioassay:** Bioactive compounds are almost always toxic at a higher dose. Thus, *in-vivo* lethality in a simple zoological organism can be used as a convenient informant for screening and fractionation in the discovery of new bioactive natural products. In the present bioactivity study all the crude extracts, VLC fractions and pure compounds showed positive results indicating that

the test samples are biologically active. Each of the test samples showed different mortality rates at different concentrations.

Plotting of log of concentration versus percent mortality for all test samples showed an approximately linear correlation. From the graphs, the median lethal concentration ( $LC_{50}$ , the concentration at which 50% mortality of brine shrimp nauplii occurred) was determined for the samples. The positive control groups showed nonlinear mortality rates at lower concentrations and linear rates at higher concentrations. There was no mortality in the negative control groups indicating the test as a valid one and the results obtained are only due to the activity of the test agents.

**RESULT AND DISCUSSION:** The brine shrimp test (BST) represents a rapid, inexpensive and simple bioassay for testing plant extract lethality which in most cases correlates reasonably well with cytotoxic and anti-tumor properties. Following the procedure of the lethality of the pet ether, ethyl acetate and chloroform fractions were determined, and the summary of the result is expressed in **Table 2**.

**TABLE 2: RESULTS OF THE TEST SAMPLES OF ALPINIA NIGRA**

Sample	$LC_{50}$ ( $\mu$ g/ml)	Regression equation	$R^2$
Vincristine sulphate (positive control)	0.563	$y = 30.056x + 56.016$	0.9168
Pet ether fraction	1.245	$y = 43.73x - 4.466$	0.945
Ethyl acetate fraction	2.151	$y = 17.71x + 11.90$	0.948
Chloroform fraction	2.737	$y = 16.60x + 4.562$	0.964

The  $LC_{50}$  values of pet ether, ethyl acetate and chloroform fraction found to be 1.245  $\mu$ g/ml, 2.151 and 2.737  $\mu$ g/ml respectively **Table 3**. The positive

control vincristine sulphate showed  $LC_{50}$  at a concentration of 0.501  $\mu$ g/ml.

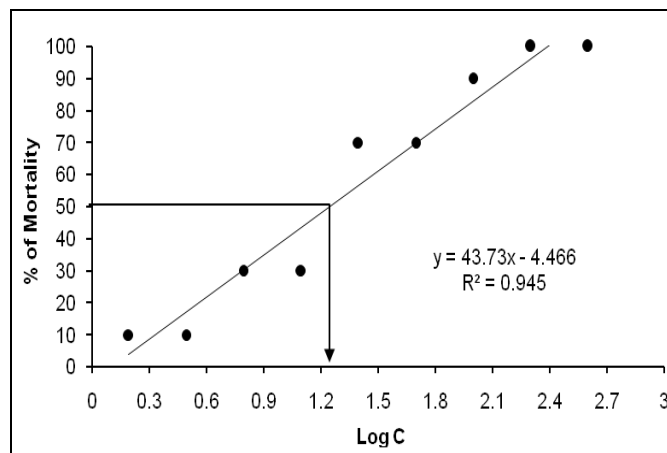
**TABLE 3: % MORTALITY OF PET ETHER, ETHYL ACETATE AND CHLOROFORM (CF) FRACTIONS OF ALPINIA NIGRA AND VINCRIStINE SULPHATE AT VARIOUS CONCENTRATIONS**

Con (C) ( $\mu$ g/ml)	Log C	% Mortality			$LC_{50}$ ( $\mu$ g/ml)			Vincristine Sulfate			
		Pet ether	Ethyl acetate	CF	Pet ether	Ethyl acetate	CF	Conc (C) ( $\mu$ g/ml)	Log C	% Mortality	$LC_{50}$ ( $\mu$ g/ml)
400	2.602	100	60	50	1.245	2.151	2.737				
200	2.301	100	50	40				20	1.301	100	
100	2	90	50	40				10	1.000	100	
50	1.699	70	40	30				5	0.698	90	
25	1.398	70	40	30				2.5	0.397	80	
12.5	1.097	30	30	20				1.25	0.096	60	
6.25	0.796	30	20	20				0.625	-0.204	50	
3.125	0.495	10	20	10				0.3125	-0.505	40	
1.563	0.194	10	20	10				0.156	-0.806	30	
0.781	-0.107	10	0	10				0.078	-1.107	20	

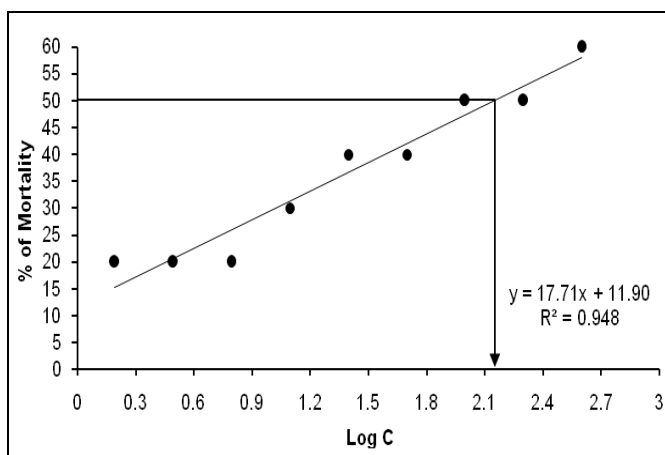
From the results of the brine shrimp lethality bioassay, it can be well predicted that the pet ether, ethyl acetate, and chloroform fractions possess cytotoxic principles. The chloroform extract was found to be considerable cytotoxic activity.

Chloroform fraction showed the highest potency ( $LC_{50}$  2.737  $\mu$ g/ml) among all the fractions.

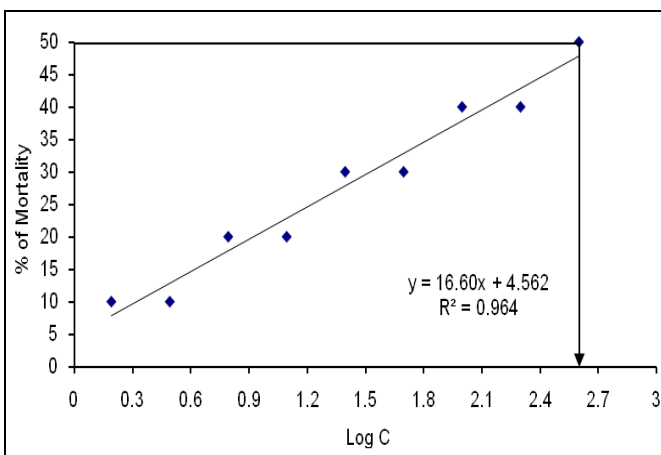
Comparison with positive control vincristine signifies that cytotoxicity exhibited by the chloroform fraction and pet ether fraction might have mild antitumor and pesticidal activity. However, this cannot be confirmed without further higher and specific tests.



**FIG. 1: EFFECT OF PET ETHER FRACTION OF RHIZOME OF *ALPINIA NIGRA* ON BRINE SHRIMP NAUPLI**



**FIG. 2: EFFECT OF ETHYL ACETATE FRACTION OF RHIZOMES OF *ALPINIA NIGRA* ON BRINE SHRIMP NAUPLII**



**FIG. 3: EFFECT OF CHLOROFORM FRACTION OF RHIZOMES OF *ALPINIA NIGRA* ON BRINE SHRIMP NAUPLII**

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**CONFLICT OF INTEREST:** Nil

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