



Received on 10 February 2014; received in revised form, 18 May 2014; accepted, 28 June 2014; published 01 July 2014

ASSESSMENT OF ANTIOXIDANT, ANTIBACTERIAL AND ANTHELMINTIC ACTIVITIES OF ETHANOL EXTRACT OF LEAVES OF *CROTALARIA PALLIDA* (AITON)

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Keywords:

Dysentery, Urinary tract infections, Vermifuge, Flavonoids

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ABSTRACT: The ethanol leaves extract of *Crotalaria pallida* (Aiton) was evaluated to investigate the Phytochemical nature and some selected pharmacological activities. The plant extract indicated the presence of combined reducing sugar, tannins, flavonoids, glycosides, alkaloids and steroids. The extract showed free radical scavenging activity with an IC₅₀ value of 37.60 µg/ml, and standard ascorbic acid showed IC₅₀ value of 16.95 µg/ml. The total phenolics content (TPC) was found to be 14 ± 0.015 mg GAE/g of dried plant extract. Total flavonoids content was found to be 20.42 ± 0.001 mg QE / 100 g of dry extract. The extract showed mild antibacterial activity against the bacterial strains *Vibrio cholera*, *Shigella flexneri*, *Shigella dysenteriae*, in comparison with standard drug kanamycin (30 µg/disc). In the anthelmintic test, the extract showed a dose-dependent decrease in paralysis time and death time of worm *Paramphistomum cervi* (trematoda) when compared with standard albendazole. These results espouse the traditional use of the leaves of this plant.

INTRODUCTION: For many years, plants are regarded as a puiissance source of natural products especially as a source of viands and therapeutic agents for a vast range of ailments¹⁻². Medicinal plants, containing versatile medicinal constituents, used in healthcare sectors and different parts of the world for a different therapeutic purpose, are a common application³. In traditional uses, the leaves of *C. pallida* (Aiton) are used as the curative agent of urinary tract infections (UTI) and vermifuge⁴. Moreover, the recent study has deciphered that it works as a potential HIV-1 protease inhibitor⁵.

Oxidation is an imperative process for living organisms for different metabolic process. Usually, a tantamount of free radicals and antiradicals are produced in a normal physiologic system but when reactive oxygen species (ROS) are increased irrationally, incur oxidative damages, provoke biochemical changes and cause diseases. For instance, diabetes mellitus, aging, arthritis, inflammation and neurodegeneration, cancer and atherosclerosis⁵⁻⁶.

Though many synthetic antioxidants have been established they are getting prohibited due to their toxic and carcinogenic comporment, and inquisition of novel antioxidant from plant parts has been started as it contains polyphenolics compounds, known properties of scavenging free radicals and can inhibit hydrolytic and oxidative enzymes⁷. Anthelmintics may also be called vermifuges or vermicides⁸.



Helminthiasis is one of the most important animal diseases, infringed upon the economy. The disease is highly rife especially in third world countries⁹. Because of the easy availability and physical resemblance of this parasite (*Paramphistomum cervi*) with the parasite of human beings, the initial evaluation of anthelmintic compounds in vitro was performed.

Plants possess many antimicrobial properties as secondary metabolites for instances, alkaloids, phenolics compounds, etc.¹⁰ The increasing case of drug-resistant of bacteria and reduced susceptibility of some strains to antibiotics raised the bacterial infections incurable and exhorted to the search for new antibacterial compounds in different plants¹¹⁻¹².

A survey of literature revealed that no research on Anti-bacterial, Anti-oxidant and Anthelmintic activity of *C. pallida* (Aiton) leaves extract was performed. For this reason, it was thought worthwhile to investigate the Anti-bacterial, Anti-oxidant and Anthelmintic activity of leaves of this plant.

MATERIALS AND METHODS:

Plant Material Collection: The leaves of *C. pallida* was collected from the Boyra, Khulna, Bangladesh in 23rd August 2012 at evening and identified by the experts of Bangladesh National Herbarium (Accession number 45236), Mirpur, Dhaka, Bangladesh. A voucher specimen has been deposited in Pharmacy Discipline, Khulna University, Khulna, Bangladesh.

Extraction: The collected leaves were separated from undesirable materials and then were washed with water. They were shade-dried for five weeks. The leaves were ground into fine powder by a suitable grinder. About 150 g of powdered material was taken in a clean glass container and soaked in 600 ml of 95% ethanol. The container was then sealed and kept for 14 days accompanying occasional stirring by a clean glass rod. The mixture then underwent a filtration process by a piece of clean, white cotton material. Then it was filtered through filter paper. The filtrate (Ethanol extract) obtained was then evaporated.

Phytochemical Screening: A small portion of the extract was taken for phytochemical screening¹³⁻¹⁴.

Test Microorganisms: Six species of both gram positive and gram negative bacteria were used for the antibacterial test. The bacterial strains were collected from the microbiology lab of Khulna University, Khulna, Bangladesh. The bacterial strains were used for the investigation are Gram-negative (*Escherichia coli*, *Shigella dysenteriae*, *Vibrio cholera*, *Shigella flexneri*) and Gram-positive (*Staphylococcus aureus*, *Streptococcus pyrogens*).

Antioxidant Assay: The free radical-scavenging activity of *C. pallida* leaves extract was evaluated by assessing its discoloration of 2, 2-diphenyl-1-picryl-hydroxyl radical (DPPH) in ethanol by a slightly modified method¹⁵. The radical scavenging activity was calculated using the following formula:

$$\text{Inhibition (\%)} = \left\{ \frac{(A-B)}{A} \right\} \times 100$$

Where, A = The absorbance of blank sample and B = The absorbance of the extract

Determination of Total Phenolics Content: The total phenolics content of *C. pallida* leaves was determined by Folin-ciocalteu assay. Briefly, a standard solution of Gallic acid (20, 40, 60, 80 µg/ml), and 0.0025 gm of the extract was dissolved in 25 ml of 80% ethanol and sonicated for 20 minutes. Then each concentration of both standard and extract was added 9 ml of distilled deionized water in each test tube. Then 1 ml of Folin-Ciocalteu was added and shaken properly. After 5 min later 10 ml of 7% of Na₂CO₃ was added. The volume was adjusted to 25ml by adding distilled water and kept in the dark for 30 min. After 30 min the UV absorbance was measured at 750 nm¹⁶.

Determination of Total Flavonoids Content: The total flavonoids content was measured with an aluminum chloride colorimetry assay. Briefly 300 µg/ml, 250 µg/ml, 200 µg/ml, 150 µg/ml, 100 µg/ml, 50 µg/ml concentration of solution of standard [Quercetin] was prepared in six test tubes. 1 ml of a solution of each test tube was taken in another six separate test tubes. 300 µg/ml concentration of the sample was prepared in another test tube. Then 4ml of distilled water was added to each test tube, and 0.3 ml of 5% NaNO₂ was added to each test tube. After 5 minutes later 0.3ml of 10% AlCl₃ was added to each test tube. 2

ml of 1M NaOH was added to each test tube. After 30 min the absorbance was taken at 510 nm wavelength¹⁶.

Anthelmintic Activity Test: The Anthelmintic activity was done on adult worm 'Paramphistoma cervi' (trematoda) due to its Physiological resemblance with the intestine parasites of human beings. Seven groups of approximately equal size of six worms in each group were used for the present study. Three groups were prepared as control (i.e. 0.2% tween-80 in water), reference i.e standard albendazole (10, 15 mg/ml) and extracts (25, 50, 100, 200 mg/ml). This test was carried out with the slight modification of the original one¹⁷. Observations were made for the time required for paralysis was noted when no movement could be observed. Death was concluded when the worms lost their motility followed with fading away of their body color.

Antibacterial Activity Test: Antibacterial activity was performed by disc diffusion method¹⁸⁻¹⁹. This method, test sample was dissolved in solvents to give the concentration of the solution to 250 µg/disc, and 500 µg/disc and kanamycin (30 µg/disc) was used as a standard. The sterile Matricel (BBL, Cockeysville, USA) filter paper discs were placed with a known amount of test samples using micropipette and dried. The disk of sample, positive control and negative control are then kept in sterile Petri dishes (120 mm in diameter) containing agar medium dipped with the test organisms using sterile transfer loop. The plates are then kept at 40 °C for facilitating maximum diffusion. The plates were then kept in an incubator for 12-18 h to allow the growth of the microorganisms. If the test material has any antibacterial activity, it will inhibit the growth of microorganism giving a clear, distinct zone called "zone of inhibition." The antibacterial activity of the test agent is determined by measuring the diameter of the zone of inhibition in term of a millimeter and compared with the standard antibiotic.

RESULTS:

Phytochemical Screening: The phytochemical analysis of *C. pallida* leaves extract revealed the presence of combined reducing sugar, steroids,

tannins, alkaloids, flavonoids, and glycosides but gums and saponins were absent.

DPPH Scavenging Activity: The leaves extract showed DPPH radical scavenging activity in a concentration-dependent manner. The IC₅₀ value for the standard and extract was measured by the following equation chronologically, $y = 44.51x - 4.718$ and $y = 41.79x - 15.83$ respectively. Where y is 50 and x is the log concentration of IC₅₀ value **Fig. 1**. The extract showed IC₅₀ value of 37.60 µg/ml which is comparable to that value of standard ascorbic acid that showed IC₅₀ value of 16.95 µg/ml. The test was done triplicates.

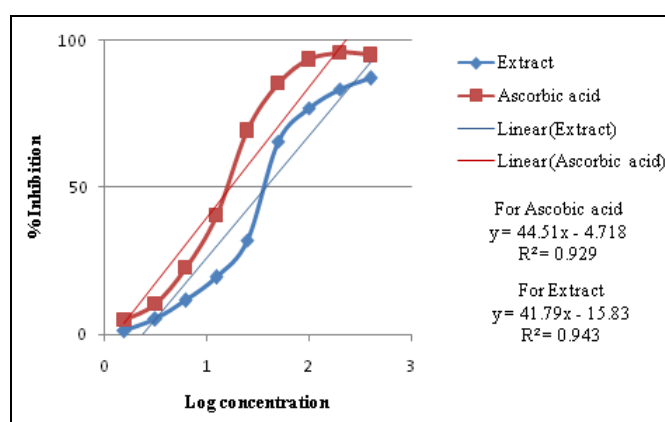


FIG. 1: DPPH SCAVENGING ASSAY (% INHIBITION vs. LOG CONC.)

Determination of Total Phenolics Content and Total Flavonoids Content: A standard curve was prepared for the determination of total phenolics content and total flavonoids content using different concentrations of standard Gallic acid and quercetin respectively. The total phenolics content of leaves extracts of *C. pallida* was expressed in term of Gallic acid equivalent. The standard curve equation: $y = 0.003x + 0.001$, $R^2 = 0.962$, **Fig. 2** where y is absorbance at 750 nm and x is total phenolics content of the extracts of *C. pallida* in mg per g²⁰. And the Total Phenolics content was found to be 14.0 ± 0.015 mg GAE/g of dried plant extract. The total flavonoids content of leaves extracts was also expressed regarding Quercetin equivalent. The standard curve equation: $y = 0.328x - 0.002$, $R^2 = 0.970$, **Fig. 3** where y is the absorbance at 510 nm, and X is the flavonoids content in mg per g²¹. The Total flavonoids content was found to be 20.42 ± 0.001 mg QE/100 g of dried plant extract.

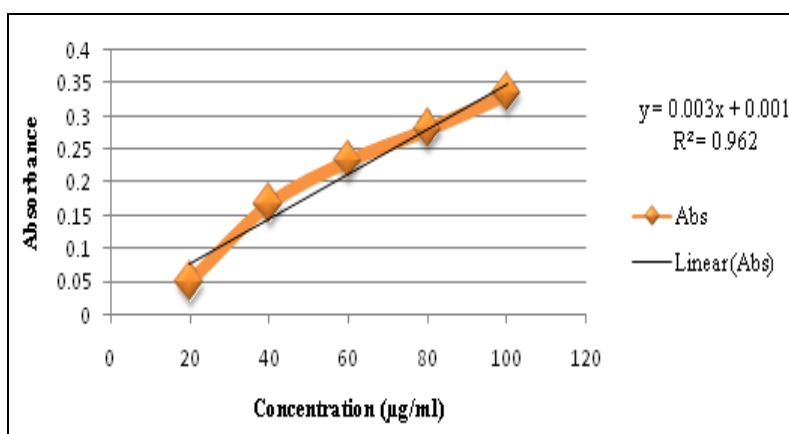


FIG. 2: LINEAR CURVE OF GALLIC ACID CONCENTRATION (µg/ml) vs. ABSORBANCE FOR DETERMINATION OF TOTAL PHENOLICS CONTENT (Abs = Absorbance)

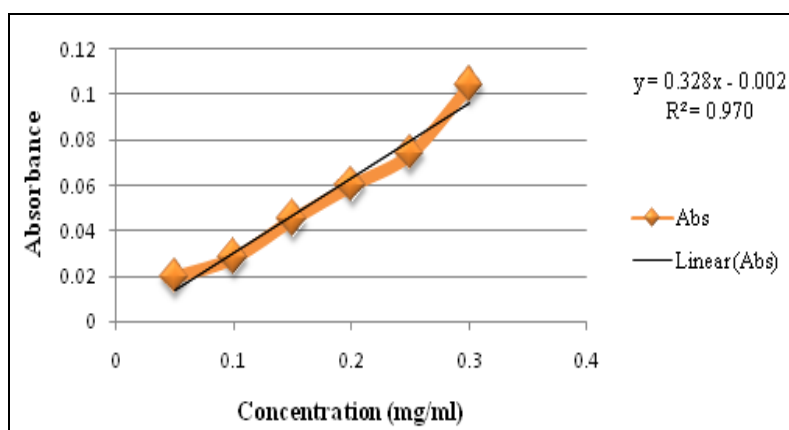


FIG. 3: LINEAR CURVE OF QUERCETIN CONCENTRATION (mg/ml) vs. ABSORBANCE FOR DETERMINATION OF TOTAL FLAVONOIDS CONTENT (Abs = Absorbance)

Antibacterial Activity: The crude extract of the leaves of *C. pallida* showed activity against the bacterial strains *Vibrio cholera*, *Shigella flexneri*, *Shigella dysenteriae*, but no activity against, *Staphylococcus aureus*, *Streptococcus pyrogens*, and *Escherichia coli*. Standard antibiotic discs of Kanamycin were used for comparison purpose.

From the results **Table 1**, the Activity Index (AI) **Table 2** were calculated using the following formula:

$$\text{Activity index (AI)} = \frac{\text{Inhibition zone of sample}}{\text{Inhibition zone of standard}}$$

TABLE 1: ANTIBACTERIAL ACTIVITY OF LEAVES EXTRACT OF *C. PALLIDA*

S. no.	Bacterial strain	Blank	Zone of inhibition		
			Kanamycin (30 µg/disc)	Extract (250 µg/disc)	Extract (500 µg/disc)
1	<i>E. coli</i>	0	19.8	0	0
2	<i>S. dysenteriae</i>	0	20.0	7.5	9.5
3	<i>S. pyrogens</i>	0	23.8	0	0
4	<i>S. aureus</i>	0	32.0	0	0
5	<i>S. flexneri</i>	0	20.0	10.5	14
6	<i>V. cholera</i>	0	24.7	9	10.5

TABLE 2: ACTIVITY INDEX OF LEAVES EXTRACT OF *C. PALLIDA*

S. no.	Bacterial strain	Extract (250 µg/disc)	Extract (500 µg/disc)
1	<i>E. coli</i>	0	0
2	<i>S. dysenteria</i>	0.38	0.48
3	<i>S. pyrogens</i>	0	0
4	<i>S. aureus</i>	0	0
5	<i>S. flexneri</i>	0.53	0.7
6	<i>V. cholera</i>	0.36	0.43

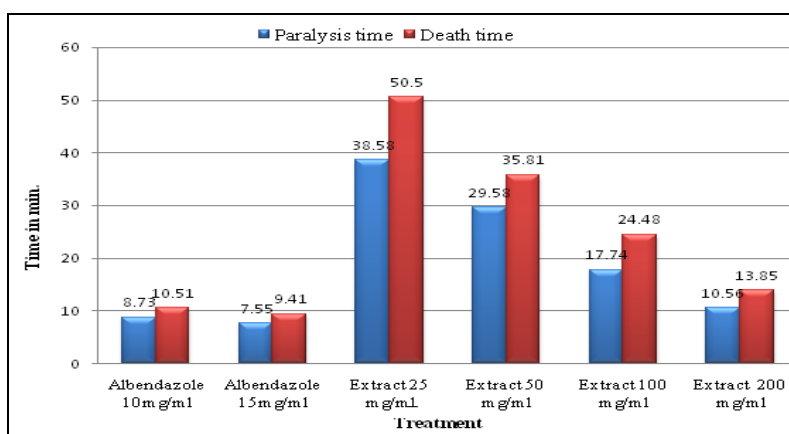


FIG. 4: ANTHELMINTIC ACTIVITY OF LEAVE EXTRACTS OF *C. PALLIDA*

TABLE 3: ANTHELMINTIC POTENCY OF ETHANOL LEAVES EXTRACT OF *C. PALLIDA*

Group	Concentration (mg/ml)	Paralysis time (min)	Death time (min)
Control	--	--	--
Standard (Albendazole)	10	8.73 ± 0.37	10.51 ± 0.39
	15	7.55 ± 0.28	9.41 ± 0.34
	25	38.58 ± 0.53	50.5 ± 0.28
Extract	50	29.58 ± 0.53	35.81 ± 0.39
	100	17.74 ± 0.46	24.48 ± 0.38
	200	10.56 ± 0.32	13.85 ± 0.49

All Values represent Mean ± SD; n=6 in each group. Comparisons made between standard versus treated groups, P<0.001 was considered significant

DISCUSSION: The presence of the bioactive plant metabolites in phytochemical screening possesses antibacterial activity. For instances, tannins are known to be helpful in treating diarrhea and dysentery.

In DPPH activity, the antioxidants were able to reduce DPPH to yellow colored diphenyl picrylhydrazone²². The method based on the reduction DPPH in alcoholic solution in the presence of a hydrogen donating antioxidant due to the formation of the nonradical form DPPH-H in the reaction. DPPH is usually used as a reagent to assess free radical and accepts an electron or hydrogen radical to become a stable diamagnetic molecule²³. The extract showed significant antioxidant activity compared to standard.

The antioxidative activities observed can be due to the different polyphenolics compounds that are, tocopherols, flavonoids and other organic acids. Many studies have shown polyphenols contribute significantly to the antioxidant activity²⁴. They act as highly effective free radical scavengers which are mainly due to their redox properties, which can play an important role in absorbing and neutralizing free radicals, quenching singlet and triplet oxygen or decomposing peroxides²⁵. The

extract showed phenolics and flavonoids content that possess a wide range of activity including antibacterial, antioxidant and anti-allergic.

The antibacterial activity of *C. pallida* probably due to the presence of flavonoids that revealed in phytochemical studies. The zone of inhibition varies within the ranges of 7.5-10.5 mm and 9.5-14 mm at the dose of 250 µg/disc and 500 µg/disc respectively. The highest zone of inhibition was found against *Shigella flexneri* (14 mm) at 500µg/disc. As it showed moderate activity against *S. dysenteriae*, *S. flexneri*, *V. cholera*, the results buttress the traditional use of this plant as a remedy of urinary disorder³, while the ethanol root extract showed moderate activity against *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Pseudomonas aeruginosa*²⁶.

The leaves extract showed significant anthelmintic activity at 200 mg/ml concentration **Table 3**.

Statistical Analysis: All measurements were repeated three times. The results are expressed as the mean values ± standard deviation. The results were statistically analyzed by ANOVA and Duncan’s multiple range tests. Statistical significance was set at p<0.001.

CONCLUSION: The present study showed that some significant phytochemical components present in the leaves extract of *C. pallida* that helps to exhibit a potent antioxidant activity and moderate antibacterial and anthelmintic activity which support the traditional uses. Further advanced inquisitions are suggested to elucidate the underlying mechanism as well as to asunder the bioactive compounds responsible for each pharmacological activity. This study has contributed to the validation of the medicinal potential of extracts of leaves of *C. pallida*.

ACKNOWLEDGEMENT: The authors are grateful to the head of the pharmacy discipline, Prof. Dr. Md. Golam Hoassain for permitting the lab facilities.

CONFLICT OF INTEREST: Nil

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How to cite this article:

Alam MH, Rahman MM, Sathi SI, Bellah SF, Mazid MA and Murshid GMM: Assessment of antioxidant, antibacterial and anthelmintic activities of ethanol extract of leaves of *Crotalaria pallida* (Aiton). Int J Pharmacognosy 2014; 1(7): 438-44. doi: 10.13040/IJPSR.0975-8232.1(7).438-44.

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