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SYNTHESIS AND BIOLOGICAL EVALUATION OF CYCLOPROPANE DERIVATIVE OF COMBRETASTATIN CA-4-PHOSPHATE

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ABSTRACT: Combretastatin A-4 (CA-4) is an anti-mitotic agent that is gaining rapid recognition among cancer biologists and clinicians as one of the newer vascular disrupting agents, (VDAs) for cancer therapy. CA-4 belongs to a group of tubulin binding natural products called combretastatin, derived from the African Bush Willow, *Combretum caffrum*. Due to CA-4's *in vivo* efficacy, a number of analogs of CA-4 have been synthesized to maximize its solubility and bioavailability. Combretastatin A-4 phosphate is a more soluble form of CA-4 that has successfully completed for treatment of acute myelogenous leukemia and relapsed ovarian cancer. This review attempts to highlight the various CA-4 analogs that have been synthesized and their effectiveness in clinical. Combretastatin A-4 phosphate (CA-4-P) is a biologically very active compound by binding to the colchicine binding site which lead to the inhibition of microtubule polymerization as well as showing antiangiogenic and anticancer effects by selectively shutting down the tumor blood flow. To avoid the disadvantage of rather low *in vivo* efficiency resulting from the isomerization of the cis stilbene derivative to the thermodynamically more stable trans-isomer, our research group started the project for CA-4 analogs synthesis. The incorporation of carbocycles with different ring sizes on the connecting carbon-bridge of the natural compound prevents the system to undergo cis-trans-isomerization. The synthesis of the cyclopropane derivative of CA-4 *via* the cyclopropanation reaction with diazomethane, and further analogs with incorporated moieties for better water solubility. This is a colorimetric assay that measures the reduction of yellow 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium bromide (MTT) by mitochondrial succinate dehydrogenase. The MTT enters the cells and passes into the mitochondria where it is reduced to an insoluble, coloured (dark purple) formazan product. The cells are then solubilised with an organic solvent (*e.g.* isopropanol) and the released, solubilised formazan reagent is measured spectrophotometrically. Since reduction of MTT can only occur in metabolically active cells the level of activity is a measure of the viability of the cells.

INTRODUCTION: Combretastatin A-4 (CA-4) (**Fig. 1**) was isolated from the bark of the South African tree *Combretum caffrum* in 1989 by Pettit and co-workers¹.

This cis-stilbene strongly inhibits tubulin polymerization by binding to the colchicine site and is found to be a cytotoxic agent against a wide variety of tumor cell lines, including multidrug-resistant lines^{2, 3}. The water-soluble sodium phosphate prodrug CA-4P (Zybrestat TM) (**Fig. 1**) is currently in several advanced clinical trials for age-related macular degeneration and anaplastic thyroid cancer based on the vascular shutdown mechanism of action^{4, 5}. Many efforts have led to the preparation of more soluble analogues⁶ for example, combretastatin CA-4 phosphate (CA-4P)

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2 or amine 3 pro-drugs ⁷. They exert antineoplastic activity through disruption of the blood flow within the tumor leading to tumor cell death ⁸. In particular, the water soluble CA- 4P has shown great clinical promise. In a phase I clinical trial, 15 patients with advanced cancer received CA-4P with bevacizumab, and this combination appeared safe and well tolerated. It was shown in this study that CA-4P induced profound vascular changes, which were maintained by the presence of bevacizumab ⁹.

Combretastatin A-4 phosphate (CA-4-P), a tubulin-depolymerising agent structurally related to colchicines, is in clinical trials as a vascular disrupting agent. Phase I trials, started in 1998, have established a maximum tolerated dose (MTD) in the range of 52–68mg/m², and showed significant changes in tumour perfusion in patients by positron emission tomography (PET) and contrast-enhanced proton magnetic resonance imaging (MRI) at these doses ¹⁰⁻¹². CA-4-P has recently entered Phase II trials in combination with conventional radiation and chemotherapy ¹³. Combretastatin A-4 was originally isolated from the South African bush willow *Combretum caffrum*. The phosphate derivative, CA-4-P, was subsequently synthesised as the more soluble prodrug ¹³. The exact mechanism of action of CA-4-P is not yet understood. *In vitro*, CA-4-P causes microtubule depolymerisation, which leads to endothelial cell shape changes ¹⁴. Other, more rapid effects include activation of the small GTPase Rho A and subsequent reorganization of the actin cytoskeleton, leading to an increase in endothelial monolayer permeability to macromolecules ¹⁵.

Further work in endothelial cells demonstrated that CA-4- P damaged mitotic spindles, arrested cells at metaphase, and led to the death of mitotic cells with a characteristic G2/M DNA content, following prolonged drug exposures ¹⁶. Mitotic arrest was associated with elevated levels of cyclin B1 protein and increased p34cdc2 activity. In animal models, cyclopropane derivative caused a prolonged and extensive drop in blood flow in tumour vessels, with much less effect in normal tissues ¹⁷. A rapid rise in tumour vascular permeability was observed leading to a catastrophic shut down of the established tumour vasculature within minutes of drug exposure, causing necrosis and secondary tumour cell death ¹⁸.

CA-4-P treatment has been shown to increase tumour hypoxia (low oxygen) within one hour of drug treatment, as measured by Eppendorf oxygen electrode or with the use of the hypoxia marker pimonidazole ¹⁹. Severe hypoxia is also known as an inherent characteristic of virtually all solid tumours, including experimental and clinical cancers, but not normal tissues. It has been correlated with poor prognosis, a more aggressive phenotype, increased metastases and resistance to therapy (see review by, ²⁰ Early *in vitro* work had analysed CA-4-P cytotoxicity under 1% oxygen conditions (mild hypoxia), showing an increase in cell kill under hypoxia ²¹. Beyond that, little is known of the effect of inherent tumour hypoxia on: cyclopropane derivative activity, or the effect of CA-4-P on gene expression. Gene regulation under hypoxia is governed by the transcription factor, hypoxia inducible factor 1 (HIF-1). HIF-1 is a heterodimer consisting of the subunits HIF-1 α and HIF-1 β /ARNT ²².

The cyclopropanation was followed by the removal of the MOM-protecting group under acidic condition, and the cyclopropyl derivative of CA-4 (1) was isolated. Subsequent derivatization of the free phenolic functionality with dibenzyl phosphite and transformation of the phosphate into the corresponding sodium salt was done to increase water solubility ²³. One of the most notable examples of such a modification can be seen in the structure of: cyclopropane derivative (**Fig. 1**), in which a disodium phosphate salt is incorporated at the phenolic position ²⁴. The delivery of combretastatin analogues to tumors *in vivo* would be challenging without modifications such as these, unless harsh organic solvents are used. These organic solvents pose their own health hazards, so the use of water soluble compounds for treatment is necessary ²⁵.

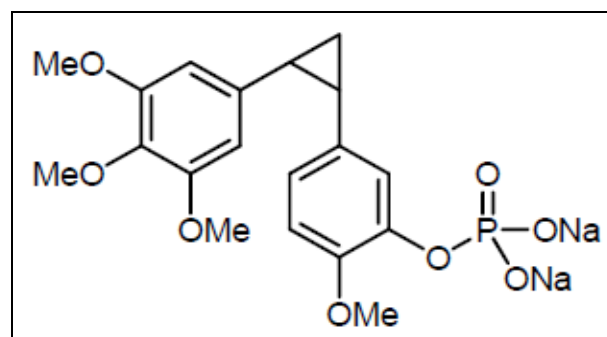
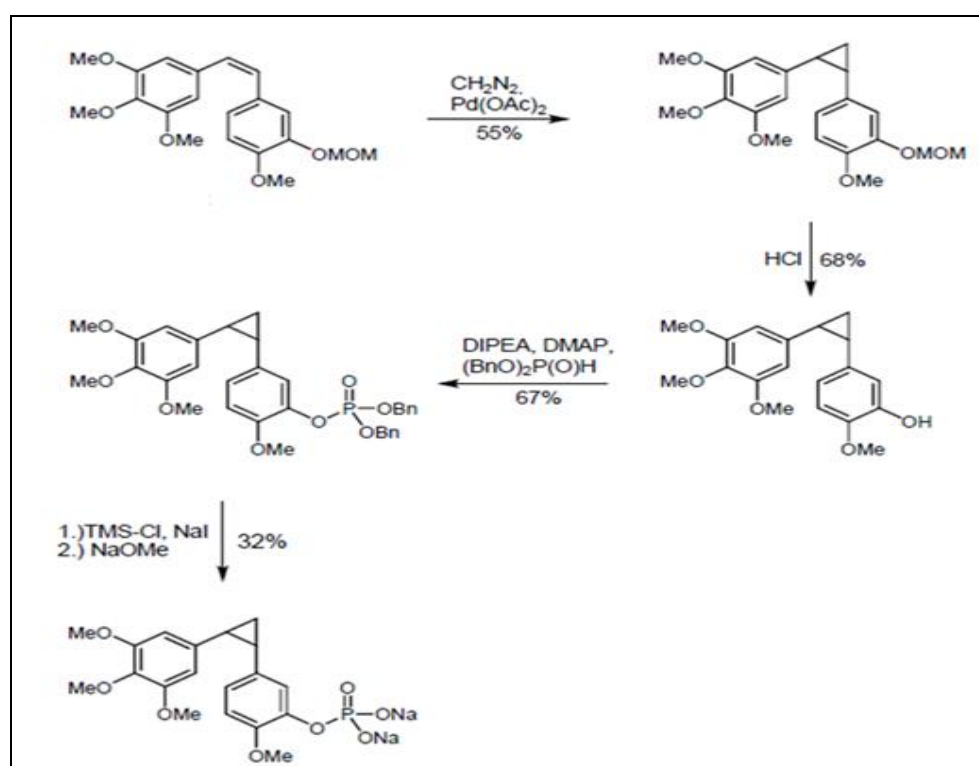


FIG. 1: CYCLOPROPANE DERIVATIVE

Further drug development entails the incorporation of bioreductive triggers in the molecular structure. While combretastatins do favor the destruction of tumor vasculature over normal blood vessels, high doses still have damaging side effects²⁶. Bioreductive triggers address this problem. Unlike other vascular disrupting agents, compounds with bioreductive trigger are introduced into the body in an inactive state. When these compounds come in contact with key bioreductive enzymes, the bioreductive trigger moiety is cleaved, leaving the active form of the compound²⁷.

As a result elongated endothelial cells round up, causing disruption of endothelial cell layer surrounding blood vessel and exposing of underlying basement membrane. This leads to blood vessel congestion and loss of blood flow, loss of oxygen and nutrient supply to tumor cells. Therefore, tumor cells undergo necrosis. In view of strong anticancer / antivascular activity exhibited by CA-4, we have synthesized some novel combretastatin analogues, and tested for their antiangiogenic activity²⁸.

Synthesis of Compounds:



SCHEME 1: SYNTHESIS OF CYCLOPROPANE DERIVATIVE

Chemicals: All the chemicals and reagents used were from CDH, New Delhi. Glass wares used were from borosil. Hydrochloric acid, Ceric Ammonium Nitrate, Dimethoxyethane, Dimethylformamide, Dimethyl sulfoxide, Diethylether, Acetic acid methylcynde *etc.*

MATERIALS AND METHODS: All compounds were purified by column chromatography and recrystallisation and confirmatory establishment of structure was done by melting point, TLC, UV, IR and ¹H NMR. Column chromatography was performed using silica gel (Qualigens, particle size 60-120mm). TLC was performed on silica gel TLC

plates. All melting points were recorded on a DECIBEL digital melting point apparatus. IR spectra were recorded on a 8400S SHIMADZU spectrometer. ¹H NMR spectra were recorded on a dpx300 spectrometer (analysis laboratory, CDRI, Lucknow).

Solvents and Chemicals Purification: The used solvents and chemicals were, if necessary, purified and dried according to common procedures as follows. Dry solvents were stored under an argon atmosphere over molecular sieve (4Å)²¹. Methylene chloride was distilled from P₂O₅, diethyl ether (Et₂O) and tetrahydrofuran (THF) were

freshly distilled from sodium / benzophenone under argon; Diisopropylamine (DIPA), diisopropyl ethylamine (DIPEA) and triethylamine (TEA), acetonitrile (MeCN), hexane and ethyl acetate were distilled from CaH₂; toluene was refluxed over sodium and freshly distilled. All other solvents were HPLC grade.

Procedure of Preparation of Compounds:

Sodium 2- methoxy- 5- (2- (3, 4, 5-trimethoxy phenyl)cyclopropyl)-phenyl phosphate: To a mixture of 267 (40mg, 0.07 mmol) and NaI (42mg, 0.28 mmol) in anhydrous acetonitrile was added TMS-Cl (35μL, 0.28 mmol) dropwise at room temperature. The reaction mixture was stirred for 3 hours, quenched with water and extracted with ethyl acetate. The organic layers were combined and the solvent was removed under reduced pressure. The residue was purified by flash chromatography (CH₂Cl₂/MeOH 5:1, 1% AcOH) affording the free phosphorous acid. The free acid was dissolved in 1mL MeOH and sodium methanolate (1.2mg, 0.023 mmol) was added. The reaction mixture was stirred for 5 min, afterwards the solvent was removed under reduced pressure affording 236 (10mg, 32%) as a white solid. R_f 0.1 (CH₂Cl₂/MeOH 3:1, 1% AcOH).

(Z)-2-Methoxy-5- (3,4,5-trimethoxystyryl)phenol (CA-4, 4): To a solution of 258 (0.14 mmol, 50mg) in 2mL methanol 0.5mL 3 M HCl were added. The reaction mixture was stirred for 24 h at room temperature. The solution was diluted with CH₂Cl₂. The organic layer was washed with water (2 x 8 mL), dried over Na₂SO₄, and the solvent was removed under reduced pressure. The residue was purified by HPLC affording 38mg (95%) of 4. 171 R_f 0.37 (hexanes/ethyl acetate 2:1);

(Z)-2-Methoxy-5-(3,4,5-trimethoxystyryl)aniline: To a solution of catalyst (Pd/CaCO₃, 0.032 mmol, 3.4mg) in 2mL ethyl acetate were added three drops of quinoline and 400μL of cyclohexene. The solution was stirred for one hour at room temperature. Alkyne 231 (100mg, 0.32 mmol) was dissolved in 1mL ethyl acetate and was added to the reaction mixture. The solution was exposed to H₂ until total consumption of the starting material (3 h). The reaction mixture was filtered and the solvent was removed under reduced pressure. The residue was purified by flash chromatography

(toluene/ethyl acetate 19:1, 1% NEt₃) affording 232 (84mg, 83%) as a yellow oil. R_f 0.27 (toluene/ethyl acetate 5:1);

Tert - butyl 2- methoxy- 5- ((3, 4, 5-trimethoxy phenyl)ethynyl)-phenylcarbamate: To a stirred solution of 293 (300mg, 0.96 mmol) in 10mL of a solvent mixture THF/H₂O in a 1:1 ratio was added K₂CO₃ (794mg, 5.76 mmol) at room temperature. After 10 min Boc₂O was added (838mg, 3.84 mmol). The reaction mixture was stirred for two days at 60 °C. The reaction was quenched with 5% aqueous HCl. The two layers were separated and the aqueous phase was extracted with ethyl acetate (3 x 15mL). The combined organic extracts were washed with water (1 x 20mL) and brine (1 x 20mL) and the solvent was removed under reduced pressure. The residue was purified by flash chromatography (hexanes/ethyl acetate 9:1), affording 262 as a colorless foam (356mg, 90%). 172 R_f 0.58 (hexanes / ethyl acetate 2:1);

Dibenzyl- 2- methoxy- 5- (2- (3, 4, 5-trimethoxy phenyl)- cyclopropyl)- phenyl phosphate: A solution of 9 (220mg, 0.67 mmol) and CCl₄ (648μL, 6.7 mmol) in 4mL acetonitrile was cooled to -10 °C and stirred for 10 min. Then DIPEA (228 μL, 1.34 mmol) and DMAP (8mg, 0.067 mmol) were added. After one minute dibenzylphosphite (221μL, 1 mmol) was added slowly and the mixture was stirred for additional 3 hours at -10 °C. After total consumption of the starting material the reaction was quenched with 0.5 M aqueous KH₂SO₄ solution. The two layers were separated and the aqueous phase was extracted with ethyl acetate (3 x 15mL). The combined organic extracts were washed with water (1 x 20mL) and brine (1x 20mL), and the solvents were removed under reduced pressure. The residue was purified by column chromatography (hexanes / ethyl acetate 3:1), affording 267 as a colorless oil (265mg, 67%). 91 R_f 0.44 (hexanes / ethyl acetate 1:1);

(Z)-Dibenzyl 2-methoxy- 5- (3, 4, 5-trimethoxy styryl) phenyl phosphate: A solution of 4 (110mg, 0.35 mmol) and CCl₄ (338μL, 3.5 mmol) in 3mL acetonitrile was cooled to -10 °C and stirred for 10 min. Then DIPEA (118μL, 0.7 mmol) and DMAP (4mg, 0.035 mmol) were added. After one minute dibenzylphosphite (116μL, 0.53 mmol) was added slowly and the mixture was stirred for an additional

3 hours at - 10 °C. After total consumption of the starting material the reaction was quenched with 0.5 M aqueous KH_2SO_4 solution. The two layers were separated and the aqueous phase was extracted with ethyl acetate (3 x 10mL). The combined organic extracts were washed with water (1 x 15mL) and brine (1 x 15mL), and the solvents were removed under reduced pressure. The residue was purified by column chromatography (hexanes/ethyl acetate 3:1), affording 259 as a colorless oil (115mg, 57%). R_f 0.44 (hexanes/ethyl acetate 1:1);

Sodium (Z)- 2- methoxy- 5- (3, 4, 5-trimethoxy styryl)phenyl phosphate: A mixture of 227 (100 mg, 0.17 mmol) and sodium iodide (99mg, 0.66 mmol) in anhydrous acetonitrile was stirred at 0 °C and TMS-Br (86 μ L, 0.66 mmol) was added dropwise. The reaction mixture was stirred for 5 min at that temperature before it was quenched with water and extracted with ethyl acetate. The combined organic layers were dried over Na_2SO_4 , filtered and the solvent was removed under reduced pressure. The residue was purified by flash column chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 3:1, 1% AcOH) affording 30mg (48%) of the free phosphorous acid. The free acid was dissolved in 1mL MeOH and sodium methanolate (8mg, 0.15 mmol) was added. The reaction mixture was stirred for 5 min, afterwards the solvent was removed under reduced pressure affording 14 (33mg, 94%) as a white solid.

Spectral Data:

Sodium 2- methoxy- 5- (2- (3, 4, 5-trimethoxy phenyl)cyclopropyl)-phenyl phosphate: ^1H NMR (400 MHz, D₂O) δ [ppm] 7.39 (s, ^1H), 6.82 (d, $J=8.56$ Hz, ^1H), 6.83 (dd, $J=8.56$, 2.05 Hz, ^1H), 6.42 (s, ^2H), 3.79 (s, ^3H), 3.73 (s, ^6H), 3.70 (s, ^3H), 2.63-2.53 (m, ^1H), 2.52-2.44 (m, ^1H), 1.53-1.44 (m, ^2H).

(Z)-2-Methoxy-5-(3,4,5-trimethoxystyryl) phenol (CA-4, 4): ^1H NMR (400 MHz, CDCl_3) δ [ppm] 6.92 (d, $J = 2.07$ Hz, ^1H), 6.80 (dd, $J = 8.26$, 2.07 Hz, ^1H), 6.73 (d, $J = 8.26$ Hz, ^1H), 6.44 (dd, $J = 12.17$, 23.67 Hz, ^2H), 5.52 (s, ^1H), 3.86 (s, ^3H), 3.84 (s, ^3H), 3.69 (s, ^6H); ^{13}C NMR (400 MHz, CDCl_3) δ [ppm] 153.01, 145.90, 145.39, 137.34, 132.84, 130.80, 129.62, 129.19, 121.25, 115.19, 110.47, 106.24, 61.06, 56.10, 56.08; IR ν 3752 3422 3152 2837 1419 1274 1005 cm^{-1} .

(Z)-2-Methoxy-5-(3,4,5-trimethoxystyryl) aniline: ^1H NMR (400 MHz, CDCl_3) δ [ppm] 6.93 (d, $J = 8.27$ Hz, ^1H), 6.90 (s, ^3H), 6.85 (dd, $J = 8.02$, 2.02 Hz, ^1H), 4.99 (s, ^2H), 3.92 (s, ^6H), 3.92 (s, ^6H), 3.91 (s, ^3H), 3.80 (s, ^3H); ^{13}C NMR (400 MHz, CDCl_3) δ [ppm] 154.14, 148.16, 139.30, 139.05, 121.16, 119.29, 117.11, 115.52, 111.73, 109.75, 90.70, 88.40, 61.35, 57.21, 56.56; IR ν 3469, 3371, 2936, 1613, 1453, 1326, 1028 cm^{-1} .

Tert-butyl 2- methoxy- 5- ((3, 4, 5-trimethoxy phenyl)ethynyl)-phenylcarbamate: ^1H NMR (400 MHz, CDCl_3) δ [ppm] 8.32 (s, ^1H), 7.16 (dd, $J = 8.34$, 2.02 Hz, ^1H), 7.09 (s, ^1H), 6.81 (d, $J = 8.34$ Hz, ^1H), 6.75 (s, ^2H), 3.89 (s, ^3H), 3.88 (s, ^6H), 3.86 (s, ^3H), 1.54 (s, ^9H); ^{13}C NMR (400 MHz, CDCl_3) δ [ppm] 153.19, 152.77, 147.69, 138.66, 128.25, 126.02, 121.09, 118.89, 116.03, 109.94, 108.83, 88.94, 88.02, 80.75, 61.11, 56.31, 55.91, 28.51; IR ν 3436, 2938, 2839, 1726, 1527, 1128 cm^{-1} .

Dibenzyl- 2- methoxy- 5- (2- (3, 4, 5-trimethoxy phenyl)- cyclopropyl)- phenyl phosphate: ^1H NMR (400 MHz, CDCl_3) δ [ppm] 7.35-7.28 (m, ^{10}H), 7.04 (s, ^1H), 6.65 (s, ^2H), 6.09 (s, ^2H), 5.16-5.05 (m, ^4H), 3.72 (s, ^3H), 3.68 (s, ^3H), 3.62 (s, ^6H), 2.41-2.29 (m, ^2H), 1.45-1.37 (m, ^1H), 1.22-1.14 (m, ^1H); ^{13}C NMR (400 MHz, CDCl_3) δ [ppm] 152.63, 148.91*, 148.86, 139.29, 139.22, 136.13, 135.95*, 135.87, 134.33, 131.33, 128.63, 128.57, 127.99, 122.83*, 122.81, 112.26, 105.0, 69.91, 69.85, 60.89, 56.05, 24.32, 23.70, 12.18; IR ν 2938, 2838, 1586, 1378, 1234, 1183, 1082 cm^{-1} .

(Z)-Dibenzyl 2- methoxy- 5- (3, 4, 5-trimethoxy styryl)phenyl phosphate: ^1H NMR (400 MHz, CDCl_3) δ [ppm] 7.38-7.27 (m, ^{10}H), 7.18-7.14 (m, ^1H), 7.07 (d, $J = 8.32$ Hz, ^1H), 6.79 (d, $J = 8.32$, Hz, ^1H), 6.49 (s, ^2H), 6.43 (dd, $J = 18.89$, 12.17 Hz, ^2H), 5.16-5.10 (m, ^4H), 3.80 (s, ^3H), 3.77 (s, ^3H), 3.68 (s, ^6H); ^{13}C NMR (400 MHz, CDCl_3) δ [ppm] 153.06, 149.93*, 149.88, 139.62*, 139.55, 137.34, 135.83, 135.76, 132.54, 130.27, 129.76, 128.63, 128.60, 128.55, 127.95, 126.65, 122.30*, 122.27, 112.39, 106.05, 69.87, 69.81, 60.94, 56.05, 56.02; IR ν 2938, 2838, 1579, 1279, 1127, 999 cm^{-1} .

Sodium (Z)- 2- methoxy- 5- (3, 4, 5-trimethoxy styryl)phenyl phosphate: ^1H NMR (free phosphorus acid, 400 MHz, MeOD) δ [ppm] 7.27 (s, ^1H), 7.01 (d, $J = 8.53$ Hz, ^1H), 6.92 (d, $J = 8.53$

Hz, ^1H), 6.55 (s, ^2H), 6.48 (dd, $J = 18.45, 12.17$ Hz, ^2H), 3.82 (s, ^3H), 3.74 (s, ^3H), 3.66 (s, ^6H).

Pharmacology: show potent cytotoxicity against M.T.T. Again, the tests were repeated with lower concentrations to determine the IC_{50} values. **Table 1** presents the IC_{50} concentration for CA-4-P (12, assay # 1107) and in comparison describes the values for the cyclopropane derivative (14, assay # 1111) and the diphosphate analog of CA-4.

Anticancer Activity by MTT Assay Method:

Procedure: Cytotoxicity testing in *in-vitro* mode will be done by using Woerdenbag *et al.*, method. Incubate 2×10^3 cells/well in the 5% CO_2 incubator for 24 h to enable them to adhere properly to the 96 well polystyrene microplates. Add test compounds, dissolved in 100% DMSO, in at least five doses, and left for 6 h. Replace the compound plus media with fresh media and incubate the cells for another 48 h in the CO_2 incubator at 37°C . Then, add $10\mu\text{l}$ MTT [3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide, and incubate the plates at 37°C for 4 h. Add $100\mu\text{l}$ DMSO to all wells and mix thoroughly to dissolve the dark blue crystals. After a few minutes at room temperature, to ensure that all crystals are dissolved, read the plates on a Spectra Max 190 Microplate Elisa reader, at 570nm. Read the plates within 1 h of adding the DMSO. Do the experiment in triplicate and calculate the inhibitory concentration (IC) values as follows:

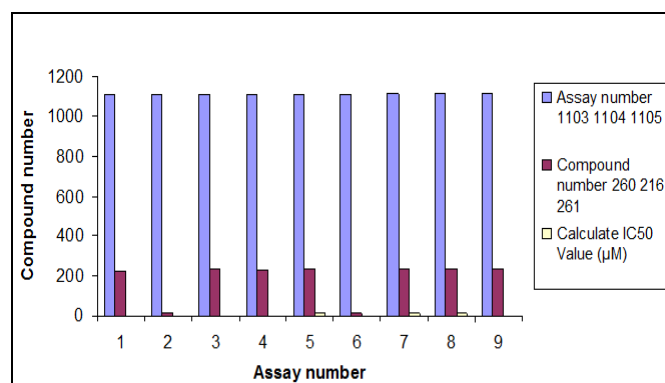
$$\% \text{ inhibition} = [1 - \text{OD} (570 \text{ nm}) \text{ of sample well} / \text{OD} (570 \text{ nm}) \text{ of control well}] \times 100.$$

IC_{50} is the concentration $\mu\text{g/ml}$ required for 50% inhibition of cell growth as compared to that of untreated control; OD = Optical density.

The same tests as described for CAM were prepared for the MTT shows the results of the first screening with an assay concentration of $30\mu\text{M}$.

RESULTS AND DISCUSSION: The concentration of the assays which were applied on the shell culture was $30\mu\text{M}$. The inhibition of shell proliferation within a certain time is investigated. The bars demonstrate that all compounds with just one exception, the alkyne derivative 261 (assay # 1105) show potent biological activity. When the inhibition of a compound is less than 50% at $30\mu\text{M}$

it is considered as inactive. It seems very interesting that also the alkyne derivatives are active at that concentration. For more detailed investigations the test was repeated with lower concentration to describe the dose-response relationship and the half maximal inhibitory concentration (IC_{50} value). The graph in Scheme 66 demonstrates the IC_{50} value for CA-4-P (12, assay # 1107) for CAM cells.



GRAPH 1: IC_{50} VALUE FOR ALL TESTED COMPOUNDS ON MTT

The result is very similar in comparison to CAM, all compounds, except compound 261 (assay # 1105), show potent cytotoxicity against M.T.T. Again, the tests were repeated with lower concentrations to determine the IC_{50} values. Presents the IC_{50} concentration for CA-4 (12, assay # 1107) and in comparison describes the values for the cyclopropane derivative (14, assay # 1111) and the diphosphate analog of CA-4 236, assay # 1114). The IC_{50} values for all tested compounds on MTT are listed in **Table 1**.

TABLE 1: IC_{50} CONCENTRATIONS, (MTT)

| Assay number | Compound number | Calculate IC_{50} Value (μM) MTT |
|--------------|-----------------|---|
| 1103 | 260 | 1.24 |
| 1104 | 216 | 20.60 |
| 1105 | 261 | >30 |
| 1106 | 226 | 0.369 |
| 1107 | 12 | 0.0034 |
| 1108 | 237 | 2.85 |
| 1109 | 227 | 1.01 |
| 1110 | 234 | 10.01 |
| 1111 | 14 | 0.202 |
| 1112 | 238 | 09.84 |
| 1113 | 235 | 08.77 |
| 1114 | 236 | 0.397 |

In this study we found that all compounds with just one exception, the alkyne derivative (261 assay # 1105) show potent biological activity. When the

inhibition of a compound is less than 50% at 30 μ M it is considered as inactive. It seems very interesting that also the alkyne derivatives are active at that concentration.

CONCLUSION: CA-4-P is a biologically very active compound by binding to the colchicine binding site which lead to the inhibition of microtubule polymerization as well as showing the Anticancer effects by selectively shutting down the tumor blood flow. To avoid the disadvantage of rather low *in vivo* efficiency resulting from the isomerization of the cisstilbene derivative to the thermodynamically more stable trans-isomer, our research group started the project for CA-4 analogs synthesis. The incorporation of carbocycles with different ring sizes on the connecting carbon-bridge of the natural compound prevents the system to undergo cis-trans-isomerization. The synthesis of the cyclopropane derivative of CA-4 (9) *via* the cyclopropanation reaction with diazomethane, and further analogs with incorporated moieties for better water solubility (268, 273) were achieved within this doctor of philosophy thesis. In cooperation with the Central Drug Research Institute, Lucknow, Uttar Pradesh. I was able to the biological activity of the compounds prepared within this work.

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CONFLICT OF INTEREST: All authors have no conflict of interest regarding this paper.

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