



Received on 26 September, 2018; received in revised form, 11 October, 2018; accepted, 23 October, 2018; published 01 November, 2018

PRELIMINARY TISSUE CULTURE STUDY OF THE CALLUS PRODUCED FROM THE AERIAL PARTS OF *ANTIGONON LEPTOPUS* (HOOK & ARN)

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Keywords:

Antigonon leptopus
Hook and Arn, vine, Polygonaceae,
HPLC, Leaf, Callus, Explant, Tissue
culture, Flavonoids, NAA

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ABSTRACT: *Antigonon leptopus* Hook & Arn (family Polygonaceae), also called: Chain of love, Queen's Wreath and Mexican creeper is a fast growing vine with stems reaching up to 20 feet long. It has heart-shaped, green leaves climbing by tendrils which wrap around many types of the supports. It is native to Mexico and commonly found in tropical Asia, Africa, the Caribbean and the Americas. It possess anti-coagulant activity, analgesic, anti-thrombin, anti-inflammatory, anti-diabetic and anti-depressant activities. Aerial parts of the vine have been used as hepatoprotective and for spleen disorders. The vine was hardly growing and greatly affected by the relatively high temperature, as well as shortage of water throughout the year in "El-Zohreya" Park, Cairo; thus to increase the plant propagation a tissue culture study was conducted. The formation of callus tissue from the leaf explants revealed promising results. Detection of the flavonoidal content by HPLC analysis was also performed on the different callus cultures, supplemented with different concentrations of the growth hormone regulator naphthalene acetic acid (NAA).

INTRODUCTION: *Antigonon leptopus*, is one of the species of belonging to the buckwheat family, Polygonaceae. It is native to Mexico, India and commonly found in tropical Asia, Africa, Caribbean and the Americas. It is known as coral vine, or Mexican creeper. It is a fast growing vine with the stems reaching to 20 feet long. It climbs by tendrils which wrap around many types of the supports. The tea prepared from the aerial portion of *A. leptopus* is used for the prevention and treatment of cough, sore throat, flu and menstrual pains. Flowers are used to treat high blood pressure. Leaves possess anti-coagulant, analgesic, anti-thrombin, anti-inflammatory and anti-diabetic activities.

Also they have been used as hepato-protective, treating asthma, liver and spleen disorders^{1, 2, 3, 4, 5, 6, 7}. Reviewing the available literature, few studies were traced on the establishment of *Antigonon leptopus* callus cultures. The vine was hardly growing and greatly affected by the relatively high temperature and by the shortage of water throughout the year in "El-Zohreya" Park, Cairo. This encouraged the author to undergo preliminary tissue culture study, with the aim of increasing the plant propagation, as well as examining the flavonoids formed in the explants produced. The study was performed on sterile Murashige and Skoog Basal Medium (MS) media, using different concentrations of growth hormone regulator naphthalene acetic acid (NAA).

MATERIALS AND METHODS:

Plant Identification: Samples of *Antigonon leptopus* used in this study were collected from "El-Zohreya" Park, Cairo, Egypt during the flowering stage (Spring 2016). A flowering branch was kindly authenticated by Mrs. Teras Labib, plant

	<p>DOI: 10.13040/IJPSR.0975-8232.IJP.5(11).700-04</p>
	<p>Article can be accessed online on: www.ijpjournal.com</p>
<p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.IJP.5(11).700-04</p>	

taxonomist of Orman garden, Giza, Egypt. The voucher specimen has been deposited in the herbarium of Department of Pharmacognosy, Faculty of Pharmacy, Cairo University “20.12.15.1”.

Plant Material (Explants): Fresh aerial parts of the vine “leaves, stems and flowers” each separately were cut into smaller sections and washed in a pyrex glass jar using detergent water mixture for about 30 min. The washed plant material was transferred to sterilizing agent solution of sodium hypochlorite 1-2% and soaked for 30 min, removed and washed with sterile distilled water to remove excess sterilizing agent.

Media: Several media were used in the tissue culture study as described in (Plant propagation by tissue culture, 2008) ⁸. The best results were obtained by using Murashige and Skoog Basal Medium (MS) ⁹. It was kindly supplied by the Agriculture Research Center, Faculty of Agriculture, Cairo University. The classic plant cell culture medium components containing macro-nutrients, micro-nutrients and vitamins are shown in **Table 1**.

TABLE 1: MURASHIGE AND SKOOG BASAL MEDIUM COMPONENTS

Components	mg/L
Macronutrients	
Potassium nitrate (KNO ₃)	1900.0
Ammonium nitrate (NH ₄ NO ₃)	1,650.0
Calcium chloride (anhydrous) (CaCl ₂)	332.20
Magnesium sulfate (anhydrous) (MgSO ₄)	180.70
Potassium phosphate monobasic (KH ₂ PO ₄)	170.0
Micronutrients	
Boric acid (H ₃ BO ₃)	6.20
Cobalt chloride hexahydrate (CoCl ₂ · 6H ₂ O)	0.0250
Cupric sulfate pentahydrate (CuSO ₄ · 5H ₂ O)	0.0250
Ferrous sulfate heptahydrate (FeSO ₄ · 7H ₂ O)	27.80
Manganese sulfate monohydrate (MnSO ₄ · H ₂ O)	16.90
Potassium iodide (KI)	0.830
Sodium molybdate dehydrate (Na ₂ MoO ₄ · 2H ₂ O)	0.250
Zinc sulfate heptahydrate (ZnSO ₄ · 7H ₂ O)	8.60
Disodium EDTA dehydrate (Na ₂ EDTA)	37.260
Carbohydrates, amino acids and vitamins	
myo-Inositol	100.0
Glycine	2.0
Nicotinic acid “Vitamin B3”	0.50
Pyridoxine hydrochloride “Vitamin B6”	0.50
Thiamine hydrochloride “Vitamin B1”	0.10

Other Media Constituents:

Growth Hormones: Auxins; naphthalene acetic acid (NAA) and cytokinins; benzylaminopurine

(BAP) to stimulate growth. They were used in different concentrations, according to the condition of the experiment applied.

- Sucrose (30 g/L) and agar (10g /L)
- NaOH 1M and HCl 1M: to adjust the pH of the media (5.8).

Sterilizing Agents:

- Detergent water mix (detergent 1 ml per litre of water).
- Bleach sterilizing solution dilute commercial bleach sodium hypochlorite (1-2%) in distilled water.
- Sterile distilled water.

Vessels and Other Material:

- Flat pyrex glass jars with plastic closures.
- Sterilized gloves, forceps and scapel blades.

All the methods adopted in the tissue culture study of the aerial parts of the vine, were based on the basic published science as described in (Cell culture basics, 2015) ^{10, 11}. This technique followed is discussed as follows:

Preparation and Sterilization of the Growing Explants: As mentioned before in plant material (explants) preparation.

Preparation and Sterilization of the Growing Medium: The commercial (MS) medium was dissolved in distilled water about 800 ml. Sucrose 30 g was added and stirred until dissolve. The pH was adjusted to (5.8) using NaOH 1M and HCl 1M as necessary while gently stirring. About 10 gm of agar was added and the volume was completed with distilled water to 1 L. The solution was heated gently while stirring until all the agar has dissolved. The still warm medium was poured into the sterilized pyrex glass jars to a depth of about 4 cm. The pyrex glass jars (with closures but not tightened) were placed in a pressure cooker and sterilized for one hour. Finally the pressure cooker was cooled, the pyrex glass jars were removed and the closures were tightened.

Transfer of Plant Aerial Parts to Tissue Culture Medium: The sterilized plant leaves, stems with attached tendrils and flowers, each separately, were cut using scalpel blades into smaller squares about

1 cm in length. Using sterile forceps, leaves were partially submerged into the medium in the sterilized pyrex glass jars. Stems and flower buds were placed so that the shoots and buds were facing upwards with the medium surface. The pyrex glass jars were tightly closed and kept in fluorescent lighted room set at 25 °C, minimum 12 h/ day, for at least 4 weeks.

Apparatus Used:

- Cell culture hood (laminar flow hood) incorporated with high efficiency particulate air (HEPA) filter: Used during the tissue culture study under aseptic conditions mentioned ¹².
- Water bath: Used for preparing the (MS) media.
- pH meter: Used for the adjustment of the pH (5.8).
- Pressure cooker: Used for the sterilization of the pyrex glass jars used.

HPLC agilent (series 1200) equipped with auto sampling injector, solvent degasser and quaternary HP pump were used. Ultraviolet (UV) detector was set at 330 nm for analysis of the flavonoidal components. Dionex bounded silica C18 column was used (4.6 × 150 mm 3 μm). Acetonitrile and 0.1% water was used as the mobile phase. Nitrogen (N₂) carrier gas was used at the injection volume of 20 μl and the rate was kept at (12 L/min).The injector temperature was set at 38 °C, while that of the detector was set at 350 °C. The range used was from 100-400 m/z. (Food Technology Research Institute, Giza, Egypt).

Induction of Callus Cultures from the Aerial Parts of *Antigonon leptopus*: Different explants were obtained from different organs of the aerial parts of *Antigonon leptopus* (leaf, petiole, stem and flowers) each alone. They were cleaned and sterilized using the sterilizing agent sodium hypochlorite solution (1-2%), as mentioned in the materials and methods section. They were germinated on sterile medium (MS) to produce seedlings from each. Germination was performed and followed up to obtain the expected callus tissue, according to the following procedure:

Dissection of uniform sized explants (of about 1cm length) from leaf, petiole, stem and flower , each separately, was performed by using sterile scalpel and forceps under aseptic conditions. The different

explants were cultured in jars containing sterile MS media, supplemented with different concentrations of various plant growth regulators as naphthalene acetic acid (NAA), indole butyric acid (IBA). In each condition twenty jars were prepared, each jar contained four explants. The culture were incubated in 16 h /day, light (1500-200 lux) at 20°C (±2 °C), over a period of 4-8 weeks. After this period, callus aging process occurred and severe browning. So, no need for the culture to be incubated over 8 weeks.

RESULTS: A photograph of the callus produced of leaf explants of *Antigonon leptopus* grown *in-vitro* on MS medium supplemented with NAA is shown in **Fig. 1**. The morphological characters of different callus cultures of leaf explants are shown in **Table 2**.

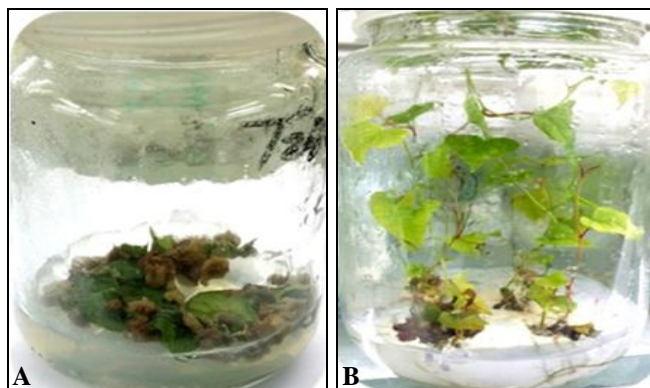


FIG. 1: PHOTOGRAPHS OF CALLUS CULTURES OF LEAF EXPLANTS OF A. LEPTOPUS GROWN IN-VITRO ON MS MEDIUM SUPPLEMENTED WITH NAA. A-4 weeks old seedling leaf explant X 0.98. B-8 weeks old seedling leaf explant X 0.9

TABLE 2: MORPHOLOGICAL CHARACTERS OF DIFFERENT CALLUS CULTURES OF LEAF EXPLANTS OF ANTIGONON LEPTOPUS

Callus cultures	Morphological characters
Leaves callus cultures on MS supplemented with (2 mg/L) NAA (Callus culture I)	Grey, compact, non-differentiated callus
Leaves callus cultures on MS supplemented with (4 mg/L) NAA (Callus culture II)	Dark grey, hollow callus showing regeneration (small shoots)
Leaves callus cultures on MS supplemented with (6 mg/L) NAA (Callus culture III)	Grey with faint brown tinge, compact callus, with small shoots and brown roots

Survival Percentage of Callus Culture Tissue of *Antigonon leptopus*: The most successful callus cultures were the following:

- Leaves callus cultures on MS supplemented with (2 mg/L) NAA.

- Leaves callus cultures on MS supplemented with (4 mg/L) NAA.
 - Leaves callus cultures on MS supplemented with (6 mg/L) NAA.
 - The other callus cultures of the explants were unsuccessful.
- The results are summarized in **Table 3**.

TABLE 3: SURVIVAL PERCENTAGE CALLUS CULTURE TISSUE OF THE LEAF EXPLANTS OF A. LEPTOPUS

Callus cultures	Survival percentage (%)
Leaves callus cultures on MS supplemented with (2 mg/L) NAA	70%
Leaves callus cultures on MS supplemented with (4 mg/L) NAA	80%
Leaves callus cultures on MS supplemented with (6 mg/L) NAA	85%

HPLC Analysis of the Flavonoid Content of Callus Cultures (I, II, III) of *Antigonon leptopus*: One gram of callus cultures (I, II, III) of the leaves were subjected to HPLC, under the adopted conditions mentioned above.

The relative concentrations of each of the detected flavonoids were determined by peak area integration. The results are recorded in **Table 4** and illustrated in **Fig. 2-5**.

TABLE 4: HPLC ANALYSIS OF THE FLAVONOID CONTENT OF CALLUS CULTURES (I, II, III) OF LEAF EXPLANTS OF *ANTIGONON LEPTOPUS*

Flavonoid	Test results of flavonoids (ppm)		
	Callus culture I	Callus culture II	Callus culture III
Luteolin-6-arabinose 8-glucoside	36.04	7.32	31.85
Luteolin-6- glucose 8-arabinoside	0.73	0.32	0.68
Apigenin-6-rhamnose 8-glucoside	1.36	0.95	4.89
Naringin	2.83	1.64	14.79
Luteolin-7- glucoside	2.73	0.46	-
Apigenin-6-glucoside 8-rhamnoside	0.76	0.94	3.48
Hesperidin	11.56	14.98	35.86
Rutin	1.26	0.48	3.13
Quercetin-3-O-glucoside	1.29	2.33	8.59
Apigenin-7-O-neohespiroside	0.37	0.72	1.97
Apigenin 7-glucoside	0.13	1.01	1.48
Kaempferol- 3,7-di-rhamnoside	0.37	1.09	3.81
Quercetrin	1.02	0.81	2.58
Quercetin	0.18	0.65	0.64
Kaempferol -3, (2-p-coumaroyl) glucose	8.11	14.99	26.92
Naringenin	0.74	1.61	1.56
Hesperitin	0.65	1.58	8.99
Kaempferol	0.33	1.40	0.65
Rhamnetin	0.36	0.81	1.02
Apigenin	0.33	0.74	1.21
Acacetin	7.86	16.09	9.15

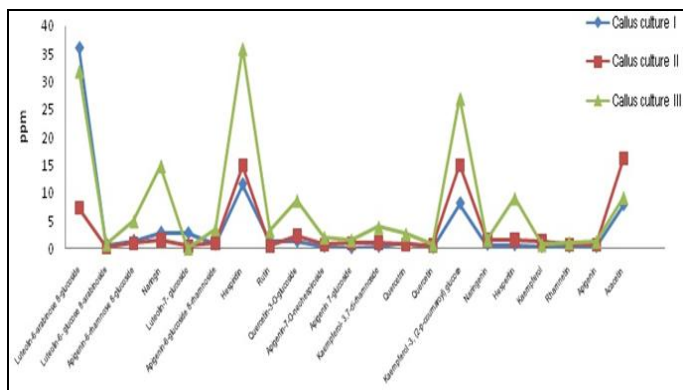


FIG. 2: LINE GRAPH SHOWING HPLC INVESTIGATION OF THE FLAVONOID CONTENT OF CALLUS CULTURES (I, II, II) OF A. LEPTOPUS

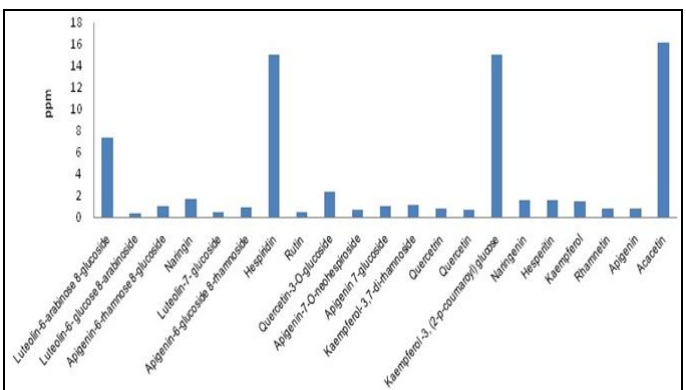


FIG. 3: HISTOGRAM SHOWING HPLC ANALYSIS OF THE FLAVONOID CONTENT OF CALLUS CULTURES (I) OF A. LEPTOPUS

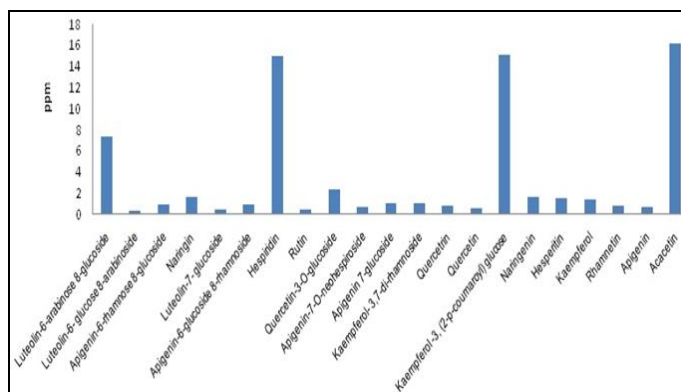


FIG. 4: HISTOGRAM SHOWING HPLC ANALYSIS OF THE FLAVONOID CONTENT OF CALLUS CULTURES (II) OF *A. LEPTOPUS*

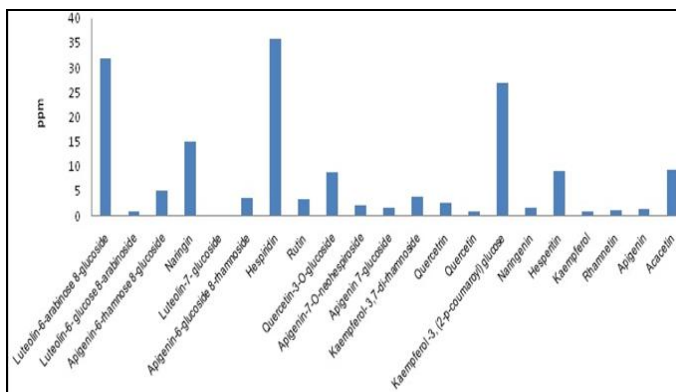


FIG. 5: HISTOGRAM SHOWING HPLC ANALYSIS OF THE FLAVONOID CONTENT OF CALLUS CULTURES (III) OF *A. LEPTOPUS*

DISCUSSION: Successful growth of leaf explants with the formation of callus was observed on the 4th week and lasted till the 8th week. Maximum callus development was achieved on MS medium supplemented with (6 mg/L) NAA (85%) (callus culture III) as shown in table (3). HPLC analysis, as shown in **Fig. 2-5**, revealed luteolin-6-arabinose-8-glucoside (36.04 ppm), acacetin (16.09 ppm) and hesperidin (35.86 ppm) as the major flavonoids in callus cultures (I, II, III) respectively. Moreover, it is obvious that callus culture III recorded the highest concentrations of flavonoidal compounds, as regards to the others. Hesperidin (35.86 ppm), luteolin-6-arabinose-8-glucoside (36.04 ppm) and kaempferol 3, (2-*p*-coumaroyl) glucose (26.92 ppm) were the major compounds in callus culture III.

CONCLUSION: The study revealed successful growth with the initiation of callus tissue. This needs further investigation, using different concentrations of growth hormones with different elicitors.

ACKNOWLEDGEMENT: The author is thankful to the Agriculture Research Center, Faculty of Agriculture, Cairo University for all the facilities provided. Special thanks are also owed to the Food Technology Research Institute, Giza, Egypt, for carrying out the HPLC analysis.

CONFLICT OF INTEREST: Nil

How to cite this article:

Ramadan NA: Preliminary tissue culture study of the callus produced from the aerial parts of *Antigonon leptopus* (Hook & Arn). Int J Pharmacognosy 2018; 5(11): 700-04. doi link: [http://dx.doi.org/10.13040/IJPSR.0975-8232.IJP.5\(11\).700-04](http://dx.doi.org/10.13040/IJPSR.0975-8232.IJP.5(11).700-04).

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