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ANTIPROLIFERATIVE ACTIVITY OF THREE WILD GROWING SPECIES IN TUNISIA: *NICOTIANA GLAUCA*, *ARTEMISIA CAMPESTRIS* AND *ASTRAGALUS GOMBO*

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ABSTRACT: Tunisian flora contains several wild plants with diverse therapeutic uses. As a contribution to the efforts to select natural sources of antitumor compounds, we investigated in this study the antiproliferative activity of several extracts of the Tunisian species: *Astragalus gombo*, *Nicotiana glauca* and *Artemisia campestris*. Dried aerial part (*A. gombo* and *A. campestris*) and leaves (*N. glauca*) were extracted with ethyl acetate, dichloromethane, and methanol. In the vitro anti-proliferative activity of the extracts was tested against the human solid tumor cell lines: HBL-100, T-47D, and WiDr. Tests were performed using the sulforhodamine B (SRB) assay. All extracts of *A. campestris* and *A. gombo* were active against all tested cell lines, with GI₅₀ values between 12 and 93 µg/ml. Dichloromethane extract of *A. campestris* was the most active extract with GI₅₀ of 12 µg/ml against HBL-100 cell line. This kind of cell seems to be the most sensitive regarding all tested extracts. Our results showed that *A. gombo*, *A. campestris* and *N. glauca* are a promising source of natural antitumor compounds. More detailed studies should be conducted especially for most active extracts.

INTRODUCTION: Cancer remains a serious problem for public health. Natural products play a highly significant role in the drug discovery and development process, particularly in the areas of cancer. With the increased resistance to the used anticancer drugs, increasing interest has been shown in research of new compounds from plants. Over 60% of these drugs were shown to be of natural origin¹.

Several species represent a significant biological potential and have been a source of active compounds for antitumor effects and cancer therapy adjuvant. Among species of Tunisian flora, several were traditionally used for various diseases.

To adapt to different stresses especially in desert zones, plants synthesize several metabolites and are thus a promising source of active molecules. These metabolites confer to these species biological activity responsible for their therapeutic use, like for *Artemisia*, *Nicotiana*, and *Astragalus* genus. Our previous studies about *Astragalus gombiformis* Pomel showed that this plant has cytotoxic, antibacterial and anticholinesterasic activities^{2, 3}. Many other investigations suggest that *Artemisia*



campestris could be beneficial for protection against diabetes and its complications, and showed that this plant exhibited antioxidant and antibacterial properties⁴⁻⁷. Other species of this genus, *Artemisia princeps*, is a potential anti-endometriotic agent that induces apoptosis of endometrial cells⁸.

Nicotiana glauca R.A. Graham, also called wild tobacco or tree tobacco, can be a source of benefit molecules such as 7-dehydrocholesterol, vitamin D₃, and other vitamin D₃-related compounds^{9,10}. *N. glauca* was used as medicinal smoke for the treatment of the ear and general skin diseases¹¹. It has a hepatoprotective effect¹². This species is toxic for animals and human can be accidentally poisoned^{13,14}. Main toxic principles of *N. glauca* are the two alkaloids nicotine and anabasine, which possess a similar structure. Others alkaloids have been identified in *N. glauca* such as anatabine, metanicotine and myosmine¹⁵⁻¹⁷. The anti-oxidant, antimicrobial and antiacetylcholinesterase activities of Tunisian *N. glauca* was recently studied by Sellem et al., (2016)¹⁸.

With many of the studies of the biological activities of plants, they remain dependent to several factors such as the natural environment of growth, the season when the plant was collected, the solvent used for extraction and the experimental conditions. We evaluate in this study the antitumor activity of three Tunisian species, *Astragalus gombo*, *N. glauca*, and *A. campestris* as a contribution to the efforts to select natural sources of anticancer compounds.

MATERIAL AND METHODS:

Plant Collection and Extracts Preparation

Samples: Aerial parts of *A. gombo* (February) have been collected on flowering stage from Djerba island in Southeast of Tunisia. Leaves of *N. glauca* and total aerial part of *A. campestris* have been collected on October from the region of Ben Gardane in Southeast of Tunisia. After air-drying in shadow, the samples have been powdered and conserved until use.

Solvents: Methanol and ethyl acetate were both from Lab-Scan. Dichloromethane and dimethyl sulphoxide (DMSO) were respectively from Carlo Erba and Sigma (St Louis, MO).

Extract Preparation: Different extracts were prepared by direct overnight maceration with organic solvents **Table 1**. Then, solvents were evaporated by rotavapor. All residues were dissolved in DMSO at 100 mg/mL concentrations as the stock solution. Both cisplatin and etoposide were used as positive control.

TABLE 1: PREPARATION OF TESTED EXTRACTS WITH DIFFERENT SOLVENT

Plant	Solvent (200 ml)	A quantity of sample (g)	Yield (%)
<i>A. campestris</i>	Methanol	10	7.18
	Dichloromethane	16	6.18
<i>A. gombo</i>	Ethyl acetate	20	6.5
	Methanol	10	7.17
<i>N. glauca</i>	Dichloromethane	20	0.99
	Ethyl acetate	20	2.82
	Methanol	7.5	11.38
	Dichloromethane	12.5	5.08
	Ethyl acetate	20	4.89

Antitumor Activity: All starting materials were commercially available research-grade chemicals and used without further purification. RPMI 1640 medium was purchased from Flow Laboratories (Irvine, UK), fetal calf serum (FCS) was from Gibco (Grand Island, NY), trichloroacetic acid (TCA) and glutamine were from Merck (Darmstadt, Germany), and penicillin G, streptomycin, DMSO and sulforhodamine B (SRB) were from Sigma (St Louis, MO).

Cells, Culture and Plating: The human solid tumor cell lines HBL-100 (breast), T-47D (breast) and WiDr (colon) were used in this study. These cell lines were a kind gift from Prof. Godefridus J. Peters (VU Medical Center, Amsterdam, The Netherlands). Cells were maintained in 25 cm² culture flasks in RPMI 1640 supplemented with 5% FBS and 2 mM L-glutamine at 37 °C, 5% CO₂, 95% humidified air incubator. Exponentially growing cells were trypsinized and resuspended in antibiotic containing medium (100 unit's penicillin G and 0.1 mg of streptomycin per ml). Single cell suspensions displaying >97% viability by trypan blue dye exclusion were subsequently counted. After counting, dilutions were made to establish the appropriate cell densities for inoculation onto 96-well microtiter plates. Cells were inoculated in a volume of 100 µl per well at densities of 10 000 (HBL-100), 15 000 (T-47D), and 20 000 (WiDr) cells per well, based on their doubling times.

Chemosensitivity Testing: Chemosensitivity tests were performed using the SRB¹⁹ assay of the NCI with slight modifications. Briefly, plant extracts were initially dissolved in DMSO at 400 times the desired final maximum test concentration of 250 µg/mL. Control cells were exposed to an equivalent concentration of DMSO (0.25% v/v, negative control). Each agent was tested in triplicate at different dilutions in the range of 2.5-250 µg/mL. The drug treatment was started on day 1 after plating.

Drug incubation times were 48 h, after which cells were precipitated with 25 µL ice-cold 50% (w/v) trichloroacetic acid and fixed for 60 min at 4 °C. The SRB assay was then performed. The optical density (OD) of each well was measured at 492 nm, using BioTek's Power Wave XS Absorbance Microplate Reader. Values were corrected for background OD from wells only containing a medium. The percentage of growth (PG) was calculated concerning untreated control cells (C) at each drug concentration level based on the difference in OD at the beginning (T_0) and end of drug exposure (T), according to NCI formulas²⁰.

Therefore, if T is greater than or equal to T_0 , the calculation is $100 \times [(T-T_0)/(C-T_0)]$. If T is lower than T_0 denoting cell killing, the calculation is 100

$\times [(T-T_0) / (T_0)]$. The effect is defined as the percentage of growth, where 50% growth inhibition (GI_{50}) represents the concentration at which PG is +50. With these calculations, a PG value of 0 corresponds to the amount of cells present at the beginning of drug exposure, while negative PG values denote net cell kill.

RESULTS AND DISCUSSION: In total, nine extracts from *N. glauca*, *A. campestris*, and *A. gombo* species were assayed for their anti-proliferative activity against three human solid tumor cell lines: HBL-100 (breast), T-47D (breast), and WiDr (colon). The results are summarized in **Table 2**. All extracts of *A. campestris* and *A. gombo* were active against all cell lines. The GI_{50} values were in the range 12-93 µg/ml. Dichloromethane extract of aerial part of *A. Campestris* showed GI_{50} of 12 µg/mL against HBL-100 cells, and it was the most active. In fact, HBL-100 seems to be the most sensitive regarding all tested extracts. *N. glauca* exhibited activity only against this cell line. For T-47D and WiDr, this species showed GI_{50} above 250 µg/mL. It is the first time, that *N. glauca* and *A. gombo* Tunisian species were tested for anti-proliferative activity against human solid tumor cell lines. However, *A. campestris* was previously studied for this activity by²¹.

TABLE 2: IN-VITRO ANTIPROLIFERATIVE ACTIVITY OF PLANT EXTRACTS AGAINST HUMAN SOLID TUMOR CELL LINES EXPRESSED AS GI_{50} VALUES GIVEN IN µg/mL

Plant	Extract	HBL-100	T-47D	WiDr
<i>A. campestris</i>	Methanol	24	45	45
	Dichloromethane	12	49	43
	Ethyl acetate	13	46	46
<i>A. gombo</i>	Methanol	49	57	42
	Dichloromethane	40	53	57
	Ethyl acetate	74	87	93
<i>N. glauca</i>	Methanol	82	>250	>250
	Dichloromethane	83	>250	>250
	Ethyle acetate	164	>250	>250
Control 1	Cisplatin	0.6	4.5	7.9
Control 2	Etoposide	0.8	12	14

In their study, the plant was collected from Beni-Khedache, a mountainous region in the Southeast of Tunisia at around 150 km from our region (Ben Gardane), which is a few km from the sea. Also, there is also a difference between tested extracts and used cells. These authors showed that *A. campestris* possess antitumor activity, with a positive correlation between this activity and the

antioxidant activity. Using HT-29 human adenocarcinoma cell line, GI_{50} values were more than 100 and 1920 µg/mL for ethanol-water and infusion extracts of aerial part, respectively. Because WiDr is considered a derivative of HT-29²², our results indicate that dichloromethane seems better solvent to extract the anti-proliferative compounds present in the aerial parts of *A.*

campestris. The essential oil was found the most active with GI₅₀ of 46.8 µg/mL²¹. Our results, in agreement with previous data, show the antiproliferative potential of *A. campestris*^{4, 5}. Ethanolic leaf extracts of *Artemisia campestris* var. *glutinosa* and *A. molinieri*, showed activity against mosquito *Culex pipiens* larvae, with low calculated lethal concentrations 50%, after 48 h of exposure²³.

Concerning *N. glauca*, few studies investigating the biological activity of this species were reported. Another Tunisian group studied the antifungal activity of *N. glauca* leaf and flower collected from the region of Monastir. Their results revealed an important antifungal effect of aqueous extracts at all concentrations tested (1, 2, 3 and 4%)²⁴. A recent study investigating *N. glauca* from Egyptian flora showed that cytotoxicity of methanol extract of flowers and leaves was 25.5, 34.2, 24.3, and 1.6 % against MCF-7, HCT-116, HepG 2, and A-549 cell lines respectively²⁵.

In our knowledge, it is the second study about the biological properties of *A. gombo* after our previous work concerning essential oil of this plant²⁶. This genus is widely studied. *Astragalus*-based herbs were known in Chinese Pharmacopoeia. This study focuses on another *Astragalus* species with antitumor effect. Recent reports were oriented to investigate the mechanism of action of *Astragalus* metabolites. Kim et al., (2013)⁸ showed that *Astragalus* polysaccharides are a potential anti-endometriotic agent. It induces apoptosis of endometrial cells by the modulation of the p38 and NF-κB pathways. *Astragalus* polysaccharides also improved experimental TNBS-induced colitis in rats through regulation of TNF-α, IL-1β and NFATc4 expression²⁷.

CONCLUSION: In the frame of selection of interesting wild Tunisian species, this study was focused on the evaluation of the antitumor potential of three wild growing plants. Extracts of *A. campestris* and *A. gombo* were active against all tested cell lines, with GI₅₀ under 100 µg/mL. Dichloromethane extract of *A. campestris* was the most active extract, with GI₅₀ equal to 12 µg/mL against HBL-100 cell line. Our results show that *A. gombo*, *A. campestris* and less *N. glauca* are a promising sources of anticancer natural

compounds. Further studies should be performed to investigate the most active extracts.

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CONFLICT OF INTEREST: Nil

REFERENCES:

1. Newman DJ, Cragg GM and Snader KM: Natural products as sources of new drugs over the period 1981-2002. *J Nat Prod* 2003; 66: 1022-1037.
2. Teyeb H, Mabrouk H, Douki W, Neffati F and Najjar MF: Anticholinesterase activity of *Astragalus gombiformis* extracts. *Journal of Biologically Active Products from Nature* 2011; 1 (5&6): 344-348.
3. Teyeb H, Zanina N, Neffati M, Douki W and Najjar MF: Cytotoxic and antibacterial activities of leaf extracts of *Astragalus gombiformis* Pomel (Fabaceae) growing wild in Tunisia. *Turk J Biol* 2012; 36: 53-58.
4. Sefi M, Fetoui H, Makni, M and Zeghal N: Mitigating effects of antioxidant properties of *Artemisia campestris* leaf extract on hyperlipidemia, advanced glycation end products and oxidative stress in alloxan-induced diabetic rats. *Food Chem Toxicol* 2010; 48: 1986-1993.
5. Akrouit A, Mighri H, Krid M, Thabet F, Turki H and Neffati M: Chemical composition and antioxidant activity of aqueous extracts of some wild medicinal plants in Southern Tunisia. *International Journal of Life Science and Medical Science* 2012; 2(1): 1-4.
6. Saoudi M, Allagui MS, Abdelmouleh A, Jamoussi K and El Feki A: Protective effects of aqueous extract of *Artemisia campestris* against pufferfish *Lagocephalus lagocephalus* extract-induced oxidative damage in rats. *Exp Toxicol Pathol* 2010; 62: 601-605.
7. Naili MB, Alghazeer RO, Saleh NA and Al-Najjar AY: Evaluation of antibacterial and antioxidant activities of *Artemisia campestris* (Asteraceae) and *Ziziphus lotus* (Rhamnaceae). *Arab J Chem* 2010; 3(2): 79-84.
8. Kim JH, Jung SH, Yang YI, Ahn JH, Cho JG, Lee KT, Baek NI and Choi JH: *Artemisia* leaf extract induces apoptosis in human endometriotic cells through regulation of the p38 and NFκB pathways. *J Pharmacol* 2013; 145: 767-775.
9. Skliar M, Curino A, Milanese L, Benassat, S and Boland R: *Nicotiana glauca*: another plant species containing vitamin D₃ metabolites. *Plant Sci* 2000; 156: 193-199.
10. Boland R, Skliar M, Curino A and Milanese L: Vitamin D compounds in plants. *Plant Sci* 2003; 164: 357-369.
11. Mohagheghzadeh A, Faridi P, Shams-Ardakani M and Ghasemi Y: Medicinal smokes. *J Ethnopharmacol* 2006; 108:161-184.
12. Janakat S and Al-Merie H: Evaluation of the hepatoprotective effect of *Pistacia lentiscus*, *Phillyrea latifolia* and *N. glauca*. *J Ethnopharmacol* 2002; 83: 135-138.
13. Steenkamp PA, Van Heerden FR and Van Wyk BE: Accidental fatal poisoning by *Nicotiana glauca*: identification of anabasine by high-performance liquid Chromatography / Photodiode Array / Mass Spectrometry. *Forensic Sci Int* 2002; 127: 208-217.

14. Plumlee KH, Holstege DM, Blanchard PC, Fiser KM and Galey FD: *Nicotiana glauca* toxicosis of cattle. J Vet Diagn Invest 1993; 5: 498-499.
15. Gaillard Y and Pepin G: Poisoning by plant material: a review of human cases and analytical determination of main toxins by High-Performance Liquid Chromatography-(tandem) Mass Spectrometry. J Chromatogr B Biomed Sci Appl 1999; 733(1-2): 181-229.
16. Botha CJ and Penrith ML: Poisonous plants of veterinary and human importance in southern Africa. Journal Ethnopharmacol 2008; 119: 549-558.
17. Saitoh F, Noma M and Kawashima N: The alkaloid contents of sixty *Nicotiana* species. Phytochemistry 1985; 24(3): 477-480.
18. Sellem I, Kaaniche F, Chakchouk Mtibaa A and Mellouli L: Anti-oxidant, antimicrobial and anti-acetyl cholinesterase activities of organic extracts from aerial parts of three Tunisian plants and correlation with polyphenols and flavonoids contents. Bangladesh J Pharmacol 2016; 11: 531-544.
19. Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D, Warren, JT, Bokesch H, Kenney S and Boyd MR: New colorimetric cytotoxicity assay for anticancer-drug screening. J Natl Cancer Inst 1990; 82: 1107-1112.
20. Monks A, Scudiero D, Skehan P, Shoemaker R, Paul K, Vistica D, Hose C, Langley J, Cronise P, Vaigro-Wolff A, Gray-Goodrich M, Campbell H, Mayo J and Boyd M: Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines. J Natl Cancer Inst 1991; 83(11): 757-766.
21. Akrouf A, Gonzalez LA, El Jani H and Madrid PC: Antioxidant and antitumor activities of *Artemisia campestris* and *Thymelaea hirsuta* from southern Tunisia. Food Chem Toxicol 2011; 49(2): 342-347.
22. Chen TR, Drabkowski D, Hay RJ, Macy M and Peterson WJ: WiDris a derivative of another colon adenocarcinoma cell line, HT-29. Can Genet Cytogenet 1987; 27: 125-34.
23. Masotti V, De Jong L, Moreau X, Rabier J, Laffont-Schwob I and Thiéry A: Larvicidal activity of extracts from *Artemisia* species against *Culex pipiens* L. mosquito: comparing endemic vs. *Ubiquist species* for effectiveness. C R Biol 2012; 335(1): 19-25.
24. Rinez A, Daami-Remadi M, Omezzine F, Ladhari A, Rinez I and Haouala R: Assessment of the antifungal activity of *Nicotiana glauca* Graham aqueous and organic extracts against some pathogenic and antagonistic fungi. Afr J Microbiol Res 2012; 6(22): 4655-4661.
25. Moustafa SMA, Menshawi BM, Wassel GA, Mahmoud K and Mounier MM: Screening of some plants in Egypt for their cytotoxicity against four human cancer cell lines. Int J Pharmtech Res 2014; 6(3): 1074-108
26. Teyeb H, Houta O, Douki W and Neffati M: Composition chimique et activité antioxydante d'huile essentielle d'*Astragalus gombo* collectée à partir de deux sites de la Tunisie. Journal de la Société Chimique de Tunisie 2012; 14(1): 63-67.
27. Yang M, Lin HB, Gong S, Chen PY, Geng LL, Zeng YM and Li DY: Effect of *Astragalus* polysaccharides on the expression of TNF- α , IL-1 β and NFATc4 in a rat model of experimental colitis. Cytokine 2014; 70: 81-86.

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